



Pharmacognostic Profile and Antibacterial Activity of Aerial Parts of *Chenopodium murale*

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Abstract

Nettleleaf goosefoot was observed on *Chenopodium murale* at Barapani, Meghalaya, India. *C. murale* is used as a leafy vegetable in the north-eastern hills region of India. This species is mainly regarded as a summer weed in the Mediterranean region. The antibacterial activity of crude petroleum ether, benzene, chloroform, acetone, ethanol and aqueous extracts of *Chenopodium murale* aerial parts were tested against *Staphylococcus capitis*, *Staphylococcus mutans*, *Pseudomonas mirabilis* and *Bacillus fragillis*. The *in vitro* antibacterial activity was performed by agar disc diffusion method. The zone of inhibition was compared with the standard drug i.e. Penicillin. Petroleum ether, chloroform, acetone and ethanol extracts were effective against the entire four test microorganism used respectively when compared to standard drug penicillin. The minimum inhibitory concentration [MIC] for *S.capitis* was 10,750,10,10,10 and 1000 mg/ml; MIC for *S.mutans* was 10,125,10,10,10 and 10 mg/ml; MIC for *P.mirabilis* was 20, 10,10,10,20 and 10 mg/ml and MIC for *B.fragillis* was 10,10,10,20,10 and 20 mg/ml for petroleum ether, benzene, chloroform, acetone, ethanol and aqueous extracts respectively suggesting the antibacterial activity of *Chenopodium murale*. Acetone extract was more effective followed by ethanol extract as antibacterial agents when compared to other extracts of aerial parts of *Chenopodium murale*. Leaves and young shoots - raw or cooked as spinach. The raw leaves should only be eaten in small quantities, see the notes above on toxicity. Seed - cooked. It can be ground into a powder and mixed with wheat or other cereals and used in making bread etc. Work is under progress to reveal the chemical nature of the active constituents responsible for the antibacterial activity.

Keywords

Aerial parts, antibacterial activity, *Chenopodium murale*, minimum inhibitory concentration, zone of inhibition.

INTRODUCTION:

Nettleleaf goosefoot was observed on *Chenopodium murale* at Barapani, Meghalaya, India. *C. murale* is used as a leafy vegetable in the north-eastern hills region of India. This species is mainly regarded as a summer weed in the Mediterranean region. However, it has spread to different geographical areas in the world including sub-tropical, temperate and cool climate regions in Canada, North America and Europe. It is generally less frequent in cooler temperatures.

The leaves and seeds of all members of this genus are more or less edible. However, many of the species in this genus contain saponins, though usually in quantities too small to do any harm. Although toxic, saponins are poorly absorbed by the body and most pass straight through without any problem. They are also broken down to a large extent in the cooking process. Saponins are found in many foods, such as some beans. Saponins are much more toxic to some creatures, such as fish, and hunting tribes have traditionally put large quantities of them in streams, lakes etc in order to stupefy or kill the fish. The plants also contain some oxalic acid, which in large quantities can lock up some of the nutrients in the food. However, even considering this, they are very nutritious vegetables in reasonable quantities. Cooking the plants will reduce their content of oxalic acid. People with a tendency to rheumatism, arthritis, gout, kidney stones or hyperacidity should take especial caution if including this plant in their diet since it can aggravate their condition.

Chenopodium murale is an ANNUAL growing to 0.6 m (2ft). It is in flower from July to October, and the seeds ripen from August to October. The species is hermaphrodite (has both male and female organs) and is pollinated by Wind. Suitable for: light (sandy), medium (loamy) and heavy (clay) soils and can grow in nutritionally poor soil. Suitable pH: mildly acid, neutral and basic (mildly alkaline) soils. It cannot grow in the shade. It prefers moist soil and can tolerate drought.

Leaves and young shoots - raw or cooked as spinach. The raw leaves should only be eaten in small quantities, see the notes above on toxicity. Seed - cooked. It can be ground into a powder and mixed with wheat or other cereals and used in making bread etc. The seed is small and fiddly; it should be soaked in water overnight and thoroughly rinsed before it is used in order to remove any saponins.

However, there is no report on antibacterial activity of this plant. In the light of the above information, the present investigation was undertaken to evaluate the antibacterial potential of different

extracts of aerial parts of *Chenopodium murale* Linn.

MATERIALS AND METHODS:**Plant Material**

Plant material used in the study consisted of aerial parts of *Chenopodium murale* Linn. was collected from the local area of Meghalaya, and authenticated by Department of Botany, Govt Maharaja College, Chhatarpur (M.P.). A voucher specimen is preserved in the Department.

Preparation of plant extract:

The dried aerial parts were coarsely powdered and subjected to successive extraction by soxhlation. The extraction was done with different solvents in their increasing order of polarity such as petroleum ether, benzene, chloroform, acetone, ethanol and distilled water. Each time the marc was dried and later extracted with other solvents. All the extract were concentrated by rotary vacuum evaporator and evaporated to dryness. 5 mg of the extract was weighed and dissolved in 5ml of DMSO which was labeled as stock 1. From stock 1 further dilution were made so as to get 10, 20, 50, 125, 250, 750 and 1000 µg/ml concentrations by using DMSO as solvent.

Microorganisms used:

All the microbial cultures, used for antimicrobial screening were procured from Microbiology Department of V.N.S. Institute of Pharmacy, Bhopal. The bacterial culture were maintained on Muller Hinton agar slants which were stored at 4°C

Antibacterial activity:**Determination of minimum inhibitory concentration (MIC)**

The extract were screened for their antibacterial activity *in vitro* by disc diffusion method [16] using *S. capitis*, *S. mutans*, *P. mirabilis* and *B. fragillis* as test organism. Agar cultures of the test microorganisms were prepared. Three to five similar colonies were selected and transferred to 5 ml broth with a loop and the broth cultures were incubated for 24 h at 37°C and suspension was checked to provide approximately 10^{10} colony forming units per ml. 0.1 ml of organism's suspension were spread evenly on the agar plates. For screening, sterile 3 mm diameter disc (Whatman filter paper No. 1) were impregnated with different concentration till saturation, dried and placed in inoculated plates of Muller Hinton agar medium. DMSO solvent was used as negative control. The plates were incubated at 37°C for 24 h. After incubation for 24 h, the results were recorded by measuring the zones of inhibition surrounding the disc and the lowest concentration of each extract which is showing inhibition of growth of bacteria was

determined as MIC. Penicillin (10 µg/ml) was used as standard for bacteria.

RESULTS AND DISCUSSION:

Microscopy of Leaf

Leaf Constant:

Sample Identity	Stomatal no.	Stomatal index	Vein islet no.	Vein termination no.	Palisade ratio
Leaves	200	9.35	200	125	9



Palisade ratio



Vein termination number

The antibacterial activity of *Chenopodium murale* aerial part extracts was studied by employing disc diffusion method against *Staphylococcus capitis*, *Staphylococcus mutans*, *Pseudomonas mirabilis* and

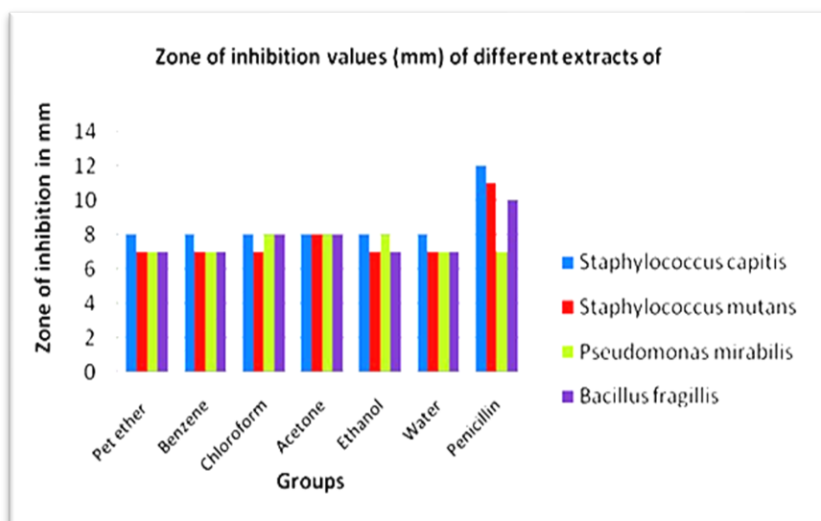
Bacillus fragillis. The results of minimum inhibitory concentration and zone of inhibition are given in Table 1 and Table 2.

Table 1: MIC values of different extracts of aerial parts of *Chenopodium murale*

Microorganism used	MIC with concentration of extract [mg/ml]					
	Petroleum ether	Benzene	Chloroform	Acetone	Ethanol	Water
<i>Staphylococcus capitis</i>	10	750	10	10	10	1000
<i>Staphylococcus mutans</i>	10	125	10	10	10	10
<i>Pseudomonas mirabilis</i>	20	10	10	10	20	10
<i>Bacillus fragillis</i>	10	10	10	20	10	20

Table 2: Zone of inhibition values (mm) of different extracts of *Chenopodium murale*

Microorganism used	Zone of inhibition (mm) of extracts and standard						
	Petroleum ether	Benzene	Chloroform	Acetone	Ethanol	Water	Penicillin
<i>Staphylococcus capitis</i>	7	6	6	7	7	7	11
<i>Staphylococcus mutans</i>	7	5	8	8	7	8	12
<i>Pseudomonas mirabilis</i>	8	7	8	8	8	7	7
<i>Bacillus fragillis</i>	9	7	7	8	9	7	11

Fig 1: Zone of inhibition [mm] of different extracts of *Chenopodium murale*


It is clear from the Table 1 and 2 and Fig 1, Petroleum ether, chloroform, acetone and ethanol extracts were effective against the entire four test microorganism used respectively when compared to standard drug penicillin. The minimum inhibitory concentration [MIC] for *S.capitis* was 10,750,10,10,10 and 1000 mg/ml; MIC for *S.mutans* was 10,125,10,10,10 and 10 mg/ml; MIC for *P.mirabilis* was 20, 10,10,10,20 and 10 mg/ml and MIC for *B.fragillis* was 10,10,10,20,10 and 20 mg/ml for petroleum ether, benzene, chloroform, acetone, ethanol and aqueous extracts respectively suggesting the antibacterial activity of *Chenopodium murale*. Work is under progress to reveal the chemical nature of the active constituents responsible for the antibacterial activity.

CONCLUSION:

The above results suggest that acetone extract was more effective followed by ethanol extract as antibacterial agents when compared to other extracts of aerial parts of *Chenopodium murale*.

REFERENCES:

- Anonymous. The Wealth of India (Raw Materials), Vol. 3. New Delhi: Publication and Information Directorate, CSIR. 2001. pp.464-69.
- P.K.Warrier. Indian Medicinal Plants- A Compendium of 500 Species, Vol. 2. Chennai: Orient Longman Pvt Ltd. 2001.pp.61-62.
- S.S.Agarwal, B.P. Yamrekar, M. Paridhavi. Clinical Useful Herbal Drug. New Delhi: Abuja Publishing House. 2005. pp.10-12.
- H.Panda. Handbook on Medicinal Herbs with Uses. New Delhi: Asia Pacific Business Press. 2005. pp.325-26.
- K.Pramila, S.Neetu, R.Anju. Medicinal plants used in traditional health care system prevalent in Western

- Himalaya. Indian J. Traditional Knowle. 2006; 5(3): 300-309.
- C.P.Khare. Indian Medicinal Plants. New Delhi: Springer International Publication. 2007. pp.141-142.
- R.P.Rastogi, B.N.Mehrotra. Compendium of Indian Medicinal Plants, Vol. 3. Reprint edn. Lucknow: CDRI. 1998. pp.162-163.
- M.Della Greca, B.D'Abrosca, A. Fiorentino, H.Previtera, A. Zarrelli. Structure elucidation and phytotoxicity of ecdysteroids from *Chenopodium murale*. Chem. Biodivers.2005; 2(4): 457-62.
- M.Della Greca, C.Di Marino, A.Zarrelli, B.D'Abrosca. Isolation and phytotoxicity of apocarotenoids from *Chenopodium murale*. J. Nat. Prod. 2004; 67(9): 1492-5.
- F.Cutillo, B.D'Abrosca, M. Della Greca, A. Zarrelli. Chenoalbicin, a novel cinnamic acid amide alkaloid from *Chenopodium murale*. Chem. Biodivers. 2004; 1(10): 1579-83.
- F.Cutillo, M.Della Greca, M. Gionti, H. Previtara, A. Zarrelli. Phenol and lignans from *Chenopodium murale*. Phytochemical Anal.2006; 17 (5): 344-9.
- D.Jhade, M.P. Padmaa, G. Usha, G. Isolation of phytoconstituents from the leaves of *Chenopodium murale* Linn. J. of Pharmacy Res. 2009; 2 (7):1192-1193.
- Y.Dai, W.C. Ye, Z.T. Wang, H. Mastuda, M. Kubo, P.P. But. Antipruritic and antinociceptive effect of *Chenopodium murale* L in mice. J. Ethnopharmacol. 2002; 81(2): 245-50.
- A.Jabbar, M.A. Zaman, Z. Labal, M. Yaseen, A. Shamin. Anthelmintic activity of *Chenopodium murale* L and *Caesalpinia crista* L against trichostrongylid nematodes of sheep. J. Ethnopharmacol. 2007; 114 (1): 86-91.
- S.Kumar, S. Biswas, D. Mandal, H.N. Roy, S. Chakraborty, S.N. Kabir. *Chenopodium murale* seed extract: a potent sperm immobilizing agent both *in vivo* and *in vitro*. Contraception. 2007; 75(1): 71-8.
- Greenwood, Slack CB, Peutherer TT. Medical Microbiology, a Guide to Microbial Infections, Pathogenesis, Immunity, Laboratory Diagnosis and Control. London: Churchill Livingstone. 2002. pp.

- "Chenopodium murale". Germplasm Resources Information Network (GRIN). Agricultural Research Service (ARS), United States Department of Agriculture (USDA). Retrieved 9 July 2013.
17. Susy Fuentes-Bazan, Pertti Uotila, Thomas Borsch: A novel phylogeny-based generic classification for *Chenopodium sensu lato*, and a tribal rearrangement of *Chenopodioideae* (*Chenopodiaceae*). In: *Willdenowia*. Vol. 42, No. 1, 2012, p. 14.
 18. BSBI List 2007 (xls). Botanical Society of Britain and Ireland. Archived from the original (xls) on 2015-06-26. Retrieved 2014-10-17.
 19. "Native American Ethnobotany" (Database). University of Michigan - Dearborn. Retrieved 27 April 2011.
 20. J. H. Maiden (1889). *The useful native plants of Australia: Including Tasmania*. Turner and Henderson, Sydney.
 21. 1990 seedlisting (1990); Tucson, Native Seed/SEARCH; pg. 17, 32
 22. Nyerges, Christopher (2017). *Foraging Washington: Finding, Identifying, and Preparing Edible Wild Plants*. "Chenopodium murale". Germplasm Resources Information Network (GRIN). Agricultural Research Service (ARS), United States Department of Agriculture (USDA). Retrieved 9 July 2013
 23. Brenan JPM, 1954. 36 *Chenopodiaceae*. In: Hutchinson J, Dalziel JM, Keay RWJ, eds. *Flora of West Tropical Africa*. Volume 1. Part 1. London, UK, Crown Agent
 24. Decary, R., 1946. *Plantes et animaux utiles de Madagascar*. Annales du Musée Colonial de Marseille, 54e année, 6e série, 4e volume, 1er et dernier fascicule. 234 pp.
 25. Mastebroek, H.D., van Soest, L.J.M. & Siemonsma, J.S., 1996. *Chenopodium L.* (grain chenopod). In: Grubben, G.J.H. & Partohardjono, S. (Editors). *Plant Resources of South-East Asia No 10. Cereals*. Backhuys Publishers, Leiden, Netherlands. pp. 79–83.
 26. Ruffo, C.K., Birnie, A. & Tengnäs, B., 2002. *Edible wild plants of Tanzania*. Technical Handbook No 27. Regional Land Management Unit/SIDA, Nairobi, Kenya. 766 pp.
 27. van Wyk, B.E. & Gericke, N., 2000. *People's plants: a guide to useful plants of southern Africa*. Briza Publications, Pretoria, South Africa. 351 pp.
 28. Aellen, P., 1959–1979. *Chenopodiaceae*. In: Rechinger, K.H. (Editor). *Hegi, G. Illustrierte Flora von Mitteleuropa*. 2nd Edition. Pteridophyta, Spermatophyta. Band 3. Angiospermae, Dicotyledones 1. Teil 2. Verlag Paul Parey, Berlin, Germany. pp. 533–747.
 29. Sathish Mohan Botsa*, Seetharam P, I. Manga Raju, Suresh P, G. Satyanarayana, Sangaraju Sambasivam, Susmitha Uppugalla, Tejeswararao D, Nanohybrid material of Co-TiO₂ and optical performance on methylene blue dye under visible light illumination. *Hybrid Advances*, 1:100008 (2022).
 30. Susmitha Uppugalla, Kavitha Rajesh, Amareswarapu V Surendra, Kiran Kumar Y, Mohammed Gayasuddin, Sriram N, Prasad P Nadedkar, Effect of *Pisonia Alba* Root Extract on Cafeteria Diet-Induced Obesity in Rats. *Journal of Pharmaceutical Negative Results*, 13, 3732–3739, 2022.
 31. N Sriram, Susmitha Uppugalla, Kavitha Rajesh, S. Someshwaran, B Senthil Kumar, Prasad P Nadedkar, Shanta K Adiki, Cognitive Enhancing and Antioxidant Activity of Ethyl Acetate Soluble Fraction of The Methanol Extract of *Pisonia Alba* Leaves in Scopalamine-Induced Amnesia. *Journal of Pharmaceutical Negative Results*, 13, 3740–3749, 2022.
 32. Brenan, J.P.M., 1954. *Chenopodiaceae*. In: Keay, R.W.J. (Editor). *Flora of West Tropical Africa*. Volume 1, part 1. 2nd Edition. Crown Agents for Oversea Governments and Administrations, London, United Kingdom. pp. 143–145.
 33. Brenan, J.P.M., 1964. *Chenopodiaceae*. In: Turrill, W.B. & Milne-Redhead, E. (Editors). *Flora of Tropical East Africa*. Crown Agents for Oversea Governments and Administrations, London, United Kingdom. 26 pp.
 34. Burkill, H.M., 1985. *The useful plants of West Tropical Africa*. 2nd Edition. Volume 1, Families A–D. Royal Botanic Gardens, Kew, Richmond, United Kingdom. 960 pp
 35. Jansen, P.C.M., 1981. Spices, condiments and medicinal plants in Ethiopia, their taxonomy and agricultural significance. *Agricultural Research Reports* 906. Centre for Agricultural Publishing and Documentation, Wageningen, Netherlands. 327 pp.
 36. Partap, T., Joshi, B.D. & Galwey, N., 1998. *Chenopods, Chenopodium spp.* Promoting the conservation and use of underutilized and neglected crops 22. International Plant Genetic Sources Institute (IPGRI), Rome, Italy. 67 pp.
 37. Rubatzky, V.E. & Yamaguchi, M., 1997. *World vegetables: principles, production and nutritive values*. 2nd Edition. Chapman & Hall, New York, United States. 843 pp.
 38. Acebes Ginovés, J.R., León Arencibia, M.C., Rodríguez Navarro, M.L., Arco Aguilar, M. del, García Gallo, A., Pérez de Paz, P.L., Rodríguez Delgado, O., Martín Osorio, V.E., Wildpret de la Torre, W., 2010. *Spermatophyta*. In: *Lista de especies silvestres de Canarias. Hongos, plantas y animales terrestres*. 2009 [ed. by Arechavaleta, M., Rodríguez, S., Zurita, N., García, A.]. Gobierno de Canarias. 122–172.
 39. Acevedo-Rodríguez, P., Strong, M.T., 2012. *Catalogue of seed plants of the West Indies*. *Smithsonian Contributions to Botany*, 98:1–1192.
 40. Aheer, G.M., Akbar, S., Chaudhri, W.M., 1997. New species of the genera *Cheletomorpha* and *Ker* (Acarina: Cheyletidae) from Pakistan. *Acarologia*, 38(2):117–121.
 41. Allam, E.K., Morsy, A.A., Ali, M.D.H., Abo-El-Ghar, A.I., 1978. Inhibitors from some higher plants inhibiting TMV and CMV infection. *Egyptian Journal of Phytopathology*, 10(1):9–20.
 42. Al-Turki, T.A., Ghafoor, A., 1996. The genus *Chenopodium L.* in Saudi Arabia. *Feddes Repertorium*, 107(3/4):189–208.
 43. Amerson, A.B., Shelton, P.C., 1976. *The natural history of Johnston Atoll*. In: *Atoll Research Bulletin No. 192*. Washington, USA: Smithsonian Institution. 479 pp.
 44. Anaya, A.L., Ramos, L., Cruz, R., Hernandez, J.G., Nava, V., 1987. Perspectives on allelopathy in Mexican

- traditional agroecosystems: a case study in Tlaxcala. *Journal of Chemical Ecology*, 13(11):2083-2101. DOI: 10.1007/BF01012873.
45. Aragón, L., Gutiérrez, W., 1992. Downy mildew on four *Chenopodium* species. (El mildiu en cuatro especies de *Chenopodium*). *Fitopatología*, 27(2):104-109.
46. Carrillo, M.M. de, Alfonso, W.P., 2003. Most important weed species in vegetable crop plantings at Quíbor valley, Lara State, Venezuela. (Especies de malezas más importantes en siembras hortícolas del valle de Quíbor, Estado Lara, Venezuela). *Bioagro*, 15(2):91-96.
47. Chandra, S.K., 2012. Invasive alien plants of Indian Himalayan region- diversity and implication. *American Journal of Plant Sciences*, 3:177-184.
48. Chaudhary, G.R., Gupta, O.P., 1991. Response of cumin (*Cuminum cyminum*) to nitrogen application, weed control and sowing methods. *Indian Journal of Agronomy*, 36(Supplement):212-216.
49. Chaudhary, S.A., Parker, C., Kasasian, L., 1981. Weeds of central, southern and eastern Arabian Peninsula. *Tropical Pest Management*, 27(2):181-190.
50. Mito T, Uesugi T, 2004. Invasive alien species in Japan: the status quo and new regulations for prevention of their adverse effects. In: *Global Environmental Research*, 8 (2) 171-191.
51. Mohamed MAA, Mohammed AA, Nasur AAH, 2018. A survey of the weed flora in garlic (*Allium sativum* L.) and onion (*Allium cepa* L.) in Dongola area, Northern State, Sudan. *International Journal of Scientific and Technology Research*. 7 (3), 28-36.
52. Uotila P, Suominen J, 1976. The *Chenopodium* species in Finland, their occurrence and means of immigration. *Annales Botanici Fennici*, 1-25.
53. USDA-NRCS, 2018. The PLANTS Database. In: The PLANTS Database. Greensboro, North Carolina, USA: National Plant Data Team. <https://plants.sc.egov.usda.gov>
54. Vafaei S H, Mahmoodi M, 2017. Presence of recombinant strain of *Cucurbit* aphid borne yellows virus in Iran. *Iranian Journal of Biotechnology*. 15 (4), 289-295. DOI:10.15171/ijb.1541
55. Walter H, 1981. Investigations into the nature and importance of weeds in sorghum in Yemen. *Plant Protection Bulletin*, Department of Plant Protection, Ministry of Agriculture, Yemen Arab Republic. 13-17.
56. Webb C J, Sykes W R, Garnock-Jones P J, 1988. *Flora of New Zealand Volume IV: Naturalized Pteridophytes, Gymnosperms, Dicotyledons*. Christchurch, New Zealand: Department of Scientific and Industrial Research. 1365 pp.
57. Wells M J, Balsinhas A A, Joffe H, Engelbrecht V M, Harding G, Stirton C H, 1986. A catalogue of problem plants in southern Africa incorporating the national weed list of South Africa. *Memoirs, Botanical Survey of South Africa*. v + 658pp.
58. Živanović B, Mitrović A, Pristov JB, Hadži-Manić KR, Čulafić L, 2010. *Chenopodium murale* L., a long-day plant as a model for physiological and biochemical research. *Biologica Nyssana*. 1 (1-2), 71-75.