



A New Simple Method Development and Validation of Ibrutinib In Bulk and Pharmaceutical Dosage Form By RP-HPLC

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Abstract

A simple, rapid, precise, accurate and sensitive reverse phase liquid chromatographic method has been developed for the determination of Ibrutinib bulk and pharmaceutical dosage form. The chromatographic method was standardized using Develosil ODS HG-5 RP C18, 5µm, 15cmx4.6mm i.d. column with UV detection at 287nm and mobile phase with the composition of 0.1% Orthophosphoric Acid: Methanol with 35:65 ratio at a flow rate of 1.0 ml/ mi. The proposed method was successfully applied to the determination of Ibrutinib in bulk and pharmaceutical dosage form. The method was linear over the range of 0-14µg/ml. The recovery was in the range of 98% to 102% and limit of detection was found to be 0.09 µg/ml and quantification was found to be 0.29 µg/ml. Different analytical performance parameters such as precision, accuracy, limit of detection, limit of quantification and robustness were determined according to International Conference on Harmonization (ICH) guidelines.

Keywords

RP-HPLC, Ibrutinib, Method development and validation, ICH Guidelines.

INTRODUCTION:

Ibrutinib is a small molecule anti-cancer drug that targets B-cell malignancies. In November 2013 Ibrutinib was approved by the FDA for the treatment of mantle cell lymphoma, and later in February 2014 for the treatment of chronic lymphocytic leukemia. Ibrutinib is also indicated for the treatment of patients with Waldenström's Macroglobulinemia (WM). As per the literature review, Ibrutinib was estimated individually by few methods like simple HPLC1, Ultra HPLC2, HPLC-MS 3method validation of ibrutinib. The objective of the work is to develop RP-HPLC method for estimation of ibrutinib in tablet dosage form with simple , rapid, accurate and

economical methods and validated for system suitability, linearity, accuracy, precision, robustness and stability of sample solution as per ICH guidelines.^[1]It is an orally administered, selective and covalent inhibitor of the enzyme Bruton's tyrosine kinase (BTK)^[3].As per the literature review, Ibrutinib was estimated individually by few methods like simple HPLC1 , Ultra HPLC2 ,HPLC-MS 3method validation of ibrutinib. The objective of the work is to develop RP-HPLC method for estimation of ibrutinib in tablet dosage form with simple, rapid, accurate and economical methods and validated for system suitability, linearity, accuracy, precision, robustness and stability of sample solution as per ICH guidelines.

The IUPAC Name of Ibrutinib is 1-[(3R)-3-[4-Amino-3-(4-phenoxyphenyl)-1Hpyrazolo [3, 4-d] pyrimidin-1-yl] piperidin-1-yl] prop-2-en-1-one.^[4]

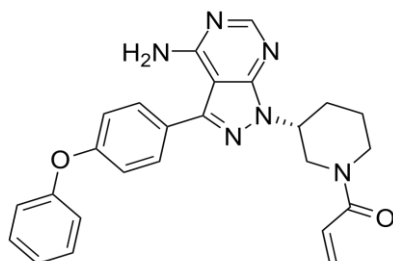


Fig 1: Chemical Structure of Ibrutinib

MATERIALS AND METHODS

HPLC Instrumentation & Conditions:

The HPLC system employed was HPLC with Empower2 Software with Isocratic with UV-Visible Detector.

Standard & sample preparation for UV-spectrophotometer analysis:

25 mg of Ibrutinib standard was transferred into 25 ml volumetric flask, dissolved & make up to volume with mobile phase. Further dilution was done by transferring 0.1 ml of the above solution into a 10ml volumetric flask and make up to volume with mobile phase. The standard & sample stock solutions were prepared separately by dissolving standard & sample in a solvent in mobile phase diluting with the same solvent. (After optimization of all conditions) for UV analysis. It scanned in the UV spectrum in the range of 200 to 400nm. This has been performed to know the maxima of Ibrutinib, so that the same wave number can be utilized in HPLC UV detector for estimating the Ibrutinib. While scanning the Ibrutinib solution we observed the maxima at 287nm. The UV spectrum has been recorded on ELICO SL-159 make UV – Vis spectrophotometer model UV-2450^[5,6]. The scanned UV spectrum is attached in the following page.

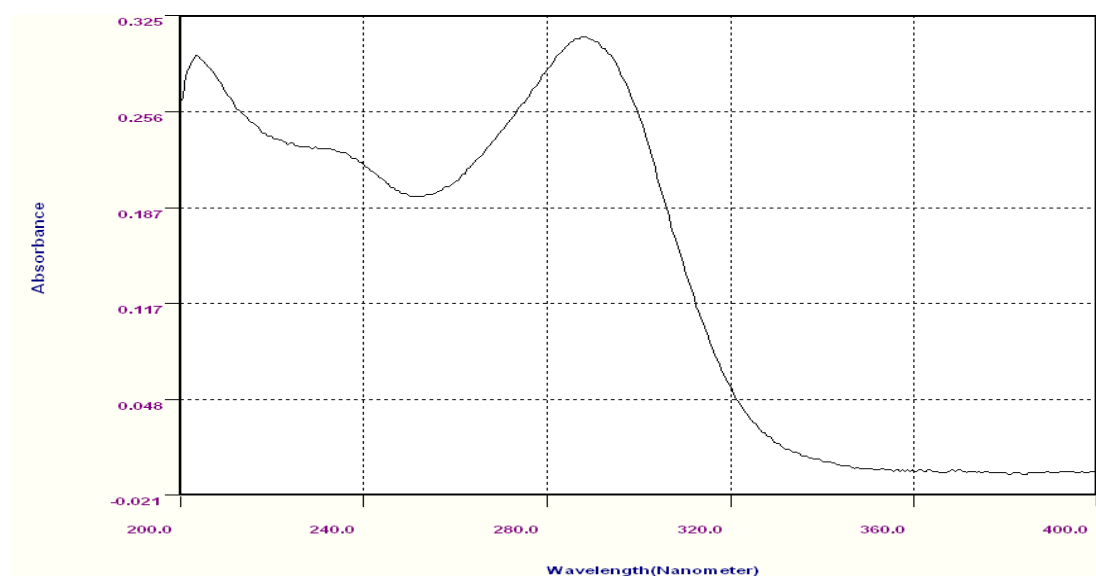


Fig 2: UV spectrum

Optimized Chromatographic Conditions:

Column: Waters ODS (C18) RP Column, 250 mm x 4.6 mm. 5 μ m.

Mobile Phase : 0.1% Orthophosphoric Acid: Methanol in the ratio 35:65(v/v)

Flow Rate: 1.0ml/minute

Wave length : 287nm

Injection volume : 20 μ l

Run time : 09mins.

Column temperature: Ambient

Sampler cooler: Ambient

MOBILE PHASE PREPARATION

Mobile phase was prepared by taking Orthophosphoric Acid: Methanol (35:65v/v). Mobile

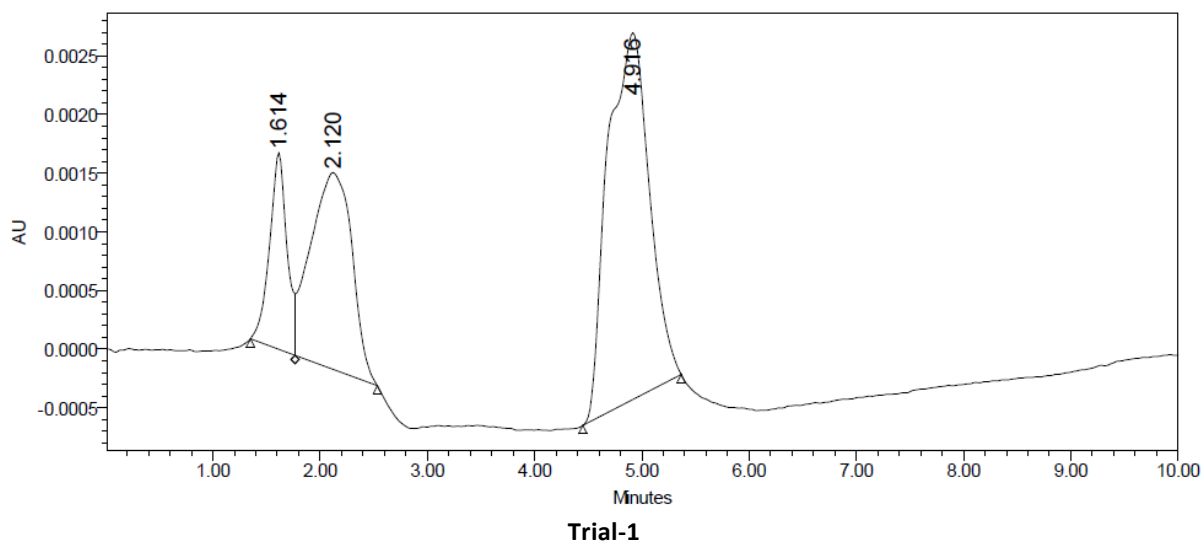
phase was filtered through 0.45 μ m membrane filter and degassed under ultrasonic bath prior to use. The mobile phase was pumped through the column at a flow rate of 1.0 ml/min.

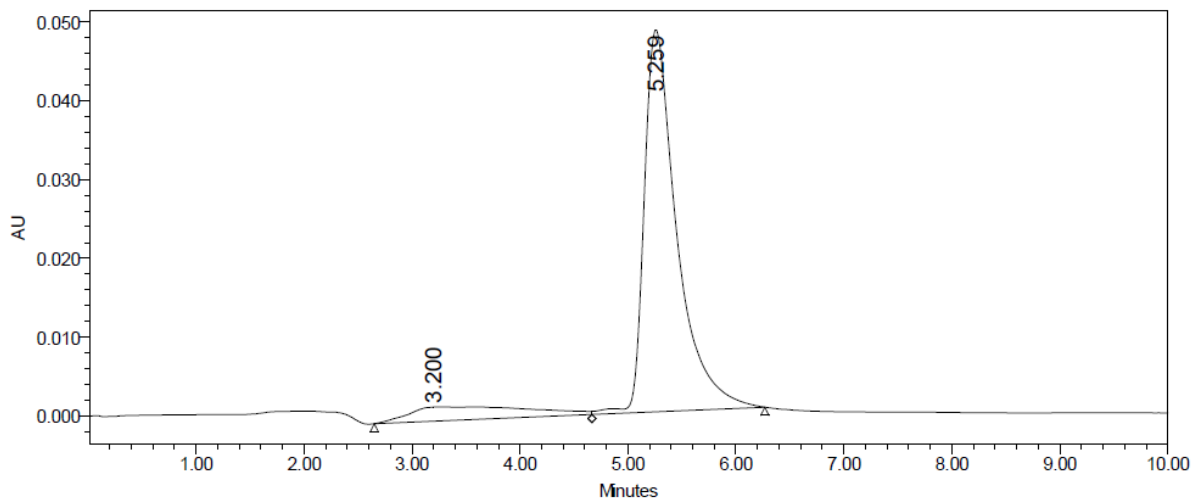
SAMPLE & STANDARD PREPARATION FOR THEANALYSIS

25 mg of Ibrutinib standard was transferred into 25 ml volumetric flask, dissolved & make up to volume with mobile phase. Further dilution was done by transferring 0.1 ml of the above solution into a 10ml volumetric flask and make up to volume with mobile phase^[7].

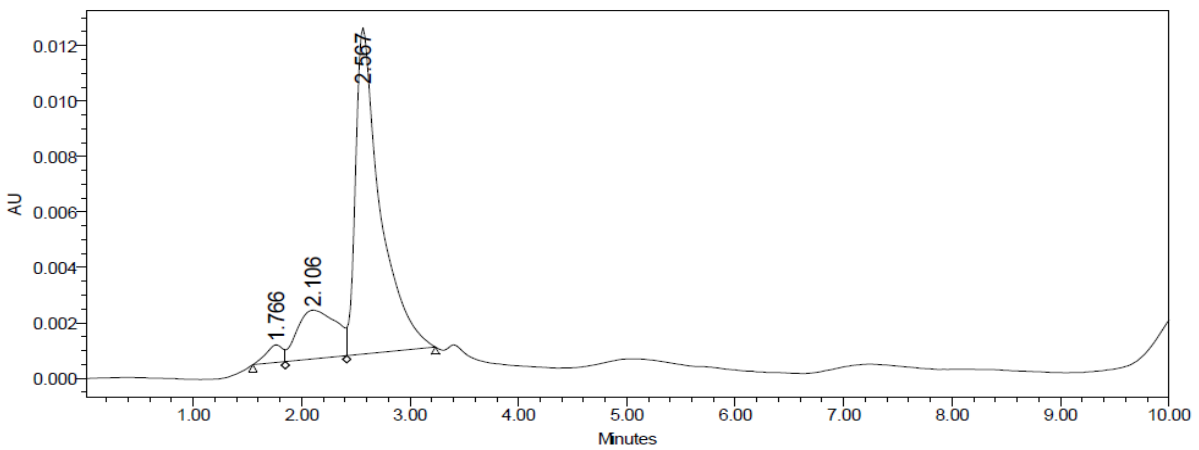
RESULT AND DISCUSSION:
Table-1: Trials for method development

Column Used	Mobile Phase	Flow Rate	Wave length	Observation	Result
Phenomenex Luna C ₁₈ , 100A, 5µm, 250mmx4.6mm i.d.	Acetonitrile: Water = 70 :30	0.8 ml/min	287nm	Peak broken at the end	Method rejected
Phenomenex Luna C ₁₈ , 100A, 5µm, 250mmx4.6mm i.d.	Methanol: Water = 60 :40	1.0 ml/min	287nm	Tailing Peaks	Method rejected
Phenomenex Luna C ₁₈ , 100A, 5µm, 250mmx4.6mm i.d.	Acetonitrile: Methanol = 70:30	1.0 ml/ min	287nm	Tailing and Frontings	Method rejected
Phenomenex Luna C ₁₈ , 100A, 5µm, 250mmx4.6mm i.d.	Acetonitrile: Methanol = 50:50	1.0 ml/ min	287nm	Tailing Peak	Method rejected
Phenomenex Luna C ₁₈ , 100A, 5µm, 250mmx4.6mm i.d.	1% Orthophosphoric Acid: Methanol = 20:80	1.0 ml/ min	287nm	Broad Peak	Method rejected
Phenomenex Luna C ₁₈ , 100A, 5µm, 250mmx4.6mm i.d.	0.1% Orthophosphoric Acid: Methanol = 35:65	1.0 ml/ min	287nm	Good Peak	Method Accepted

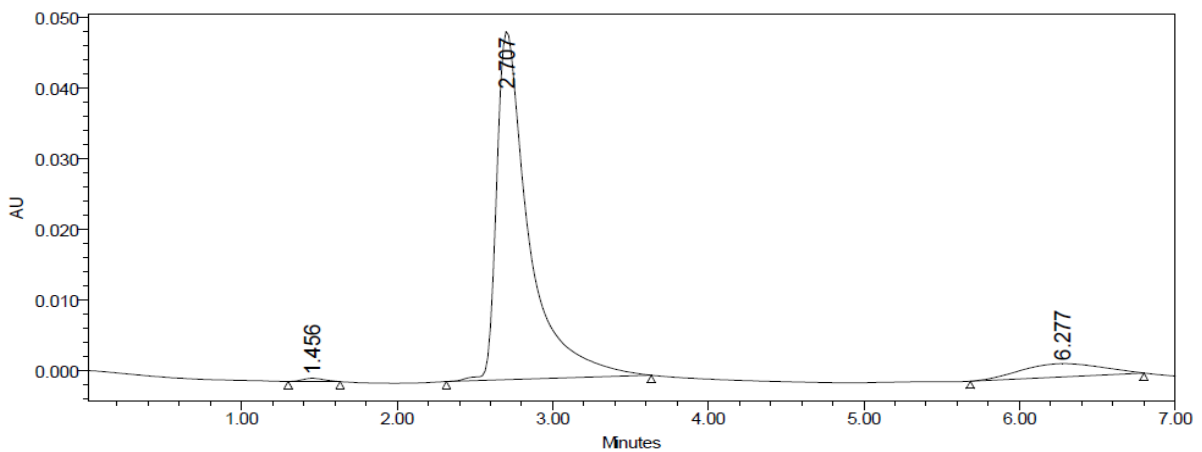




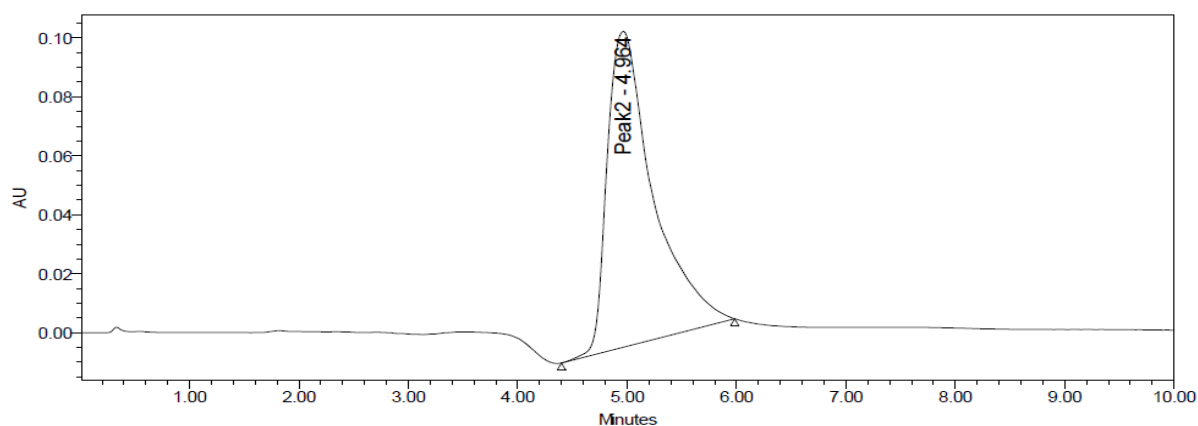
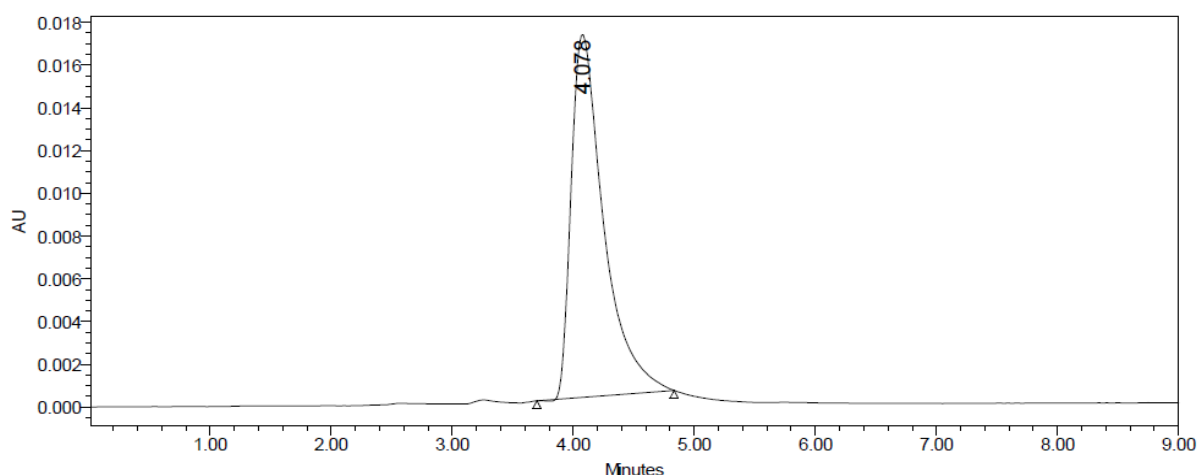
Trial-2



Trial-3



Trial-4


Trial-5

Trial-6
Table 2: Peak results

Rt	Peak Area	Theoretical Plates	Tailing Factor
4.078	327368	4687	1.29

METHOD VALIDATION:
Accuracy: Recovery study:

To determine the accuracy of the projected technique, recovery studies were distributed by adding totally different amounts (80%, 100%, and

120%) of pure drug of Ibrutinib were taken and side to the pre-analysed formulation of concentration 10 μ g/ml^[8,9]. From that proportion recovery values were calculated.

Table-3: Accuracy Readings

Sample ID	Concentration (μ g/ml)			%Recovery of Pure drug	Statistical Analysis
	Conc. Injected	Conc. Recovered	Peak Area		
S ₁ : 80 %	8	7.991	269458	99.887	Mean= 100.4953 S.D. = 0.701041 % R.S.D.=0.697586
S ₂ : 80 %	8	8.101	273103	101.262	
S ₃ : 80 %	8	8.027	270641	100.337	
S ₄ : 100 %	10	9.914	332711	99.14	Mean= 99.22667 S.D. = 0.185831 % R.S.D.= 0.18728
S ₅ : 100 %	10	9.910	332567	99.10	
S ₆ : 100 %	10	9.949	333854	99.44	
S ₇ : 120 %	12	11.939	399277	99.491	Mean= 99.894 S.D. = 0.786211 % R.S.D.=0.787045
S ₈ : 120 %	12	11.927	398886	99.391	
S ₉ : 120 %	12	12.096	404441	100.80	

Precision:
Repeatability

The precision of each method was ascertained separately from the peak areas & retention times

obtained by actual determination of six replicates of a fixed amount of drug. Ibrutinib (API) the percent relative standard deviations were calculated for Ibrutinib^[10].

Table-4: Repeatability Results of Precision

HPLC Injection Replicates of Leucovorin	Retention Time	Peak Area
Replicate – 1	4.399	1067796
Replicate – 2	4.399	1073916
Replicate – 3	4.398	1077381
Replicate – 4	4.392	1064998
Replicate – 5	4.392	1060633
Replicate –6	4.393	1061645
Average		1067728
Standard Deviation		6727.127
% RSD		0.630041

Intra day & Inter day: The intra & inter day variation of the method was carried out & the high values of mean assay & low values of standard deviation & %

RSD (% RSD < 2%) within a day & day to day variations for Ibrutinib revealed that the proposed method is precise^[11,12].

Table-5: Results of Intra day&Inter day

Conc. Of Ibrutinib (API) (µg/ml)	Observed Conc. Of Ibrutinib (µg/ml) by the proposed method			
	Intra day		Inter day	
	Mean (n=6)	% RSD	Mean (n=6)	% RSD
8	7.96	1.09	8.06	1.06
10	10.09	0.95	9.86	0.92
12	12.03	0.96	11.96	0.99

Linearity and Range

The calibration curve showed good linearity in the range of 0-14µg/ml, for Ibrutinib (API) with

correlation coefficient (r^2) of 0.998 (Fig-37). A typical calibration curve has the regression equation of $y = 32880x + 6712$ for Ibrutinib^[13].

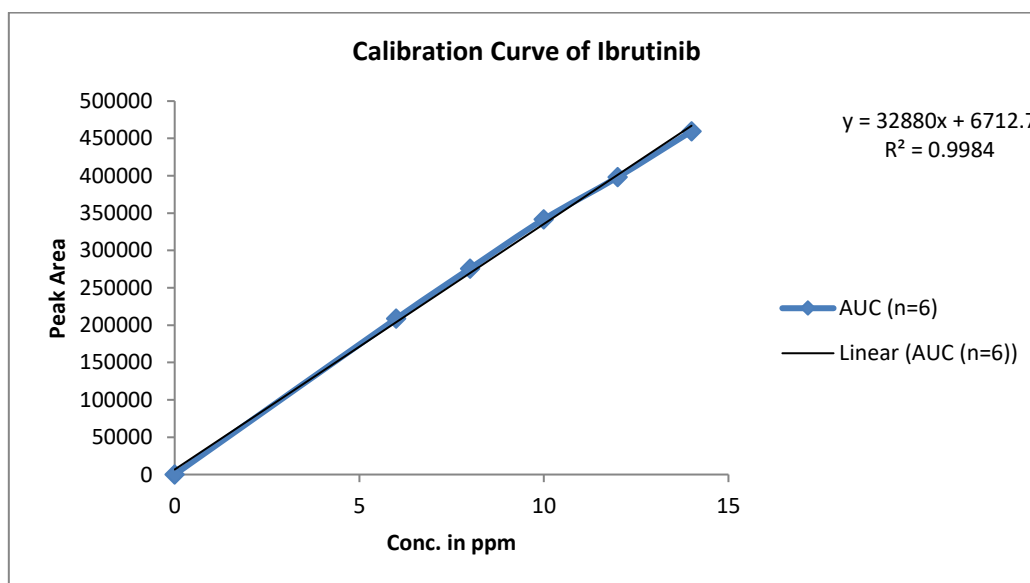

Fig-3: Calibration curve of Ibrutinib (API)

Table-6: Linearity Results of Ibrutinib

CONC.	AUC (n=6)
0	0
6	208757
8	275909
10	341782
12	398192
14	459633

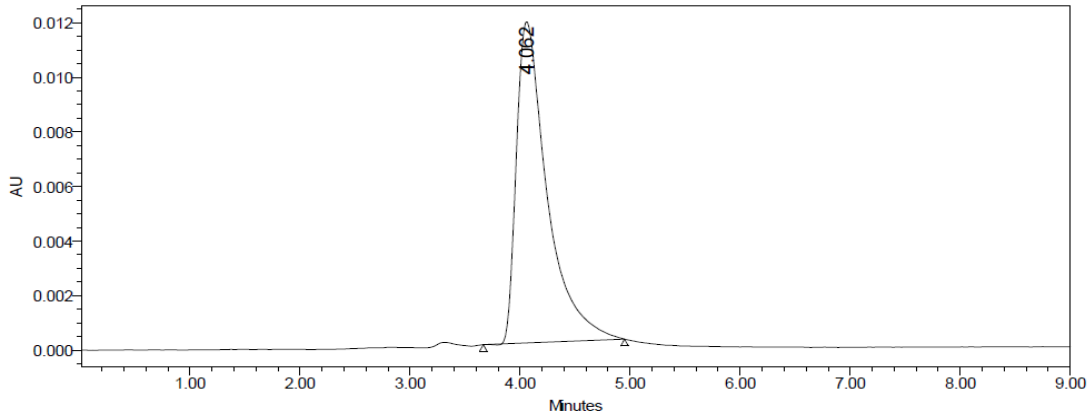


Fig 4: Calibration of Ibrutinib concentration in 6 ppm

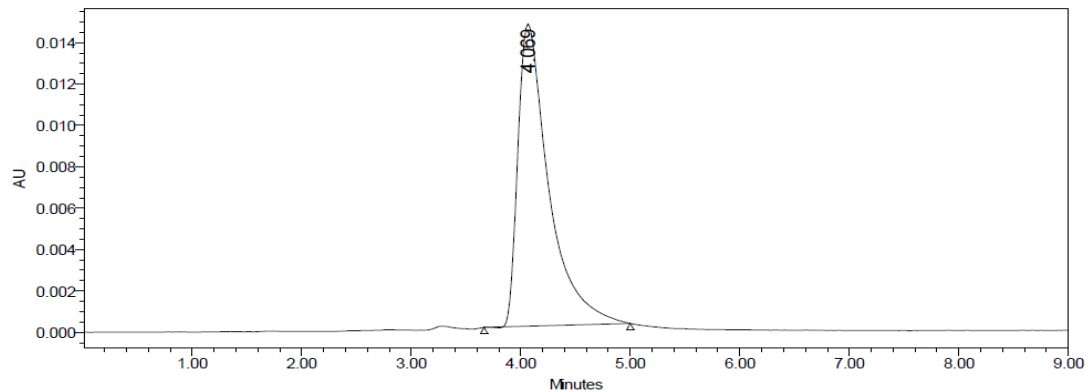


Fig 5: Calibration of Ibrutinib concentration in 8 ppm

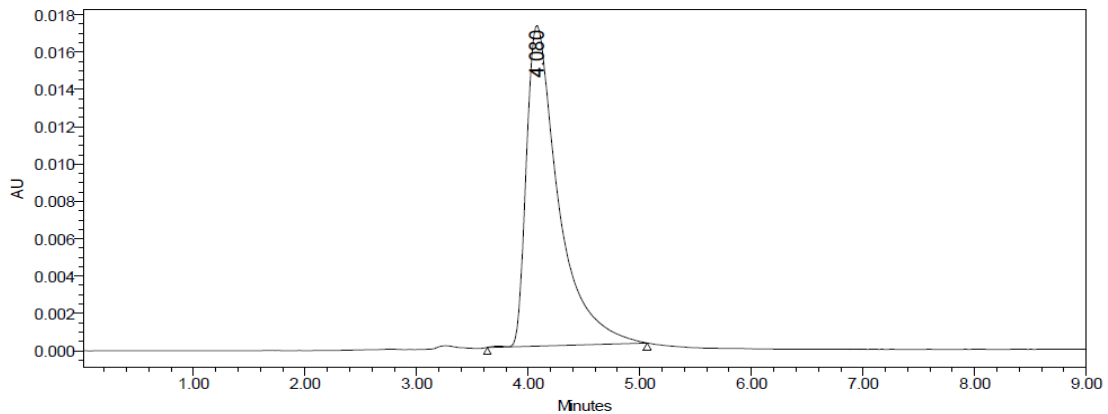


Fig 6: Calibration of Ibrutinib concentration in 10 ppm

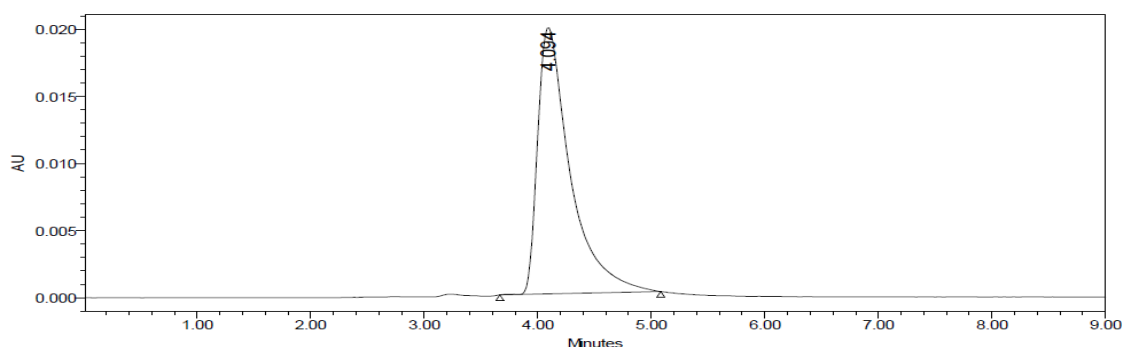


Fig 7: Calibration of Ibrutinib concentration in 12 ppm

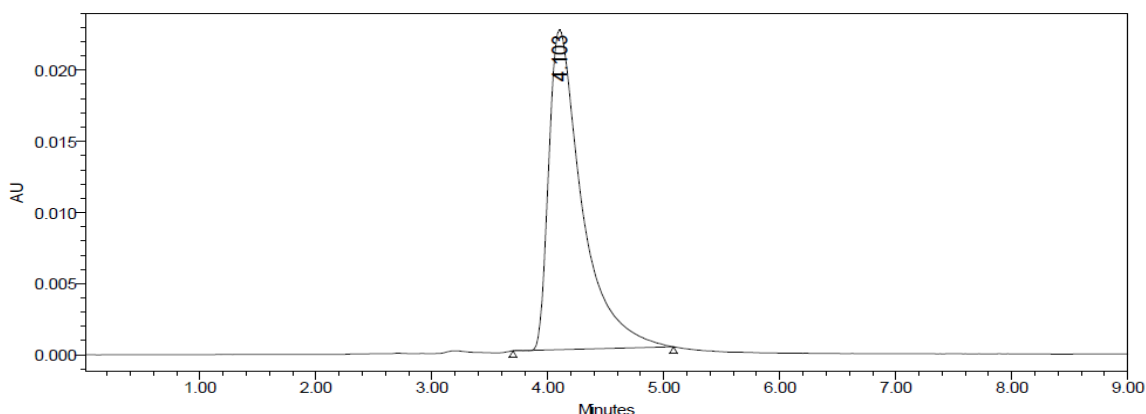


Fig 8: Calibration of Ibrutinib concentration in 14 ppm

LOD & LOQ: The Minimum concentration level at which the analyte can be reliably detected (LOD) & quantified (LOQ) were found to be 0.09 & 0.29 $\mu\text{g/ml}$ respectively.

System Suitability Parameter

System quality testing is an integral part of several analytical procedures. The tests area

unit supported the construct that the instrumentation, physics, Associate in Nursing analytical operations and samples to be analysed represent an integral system that may be evaluated intrinsically [14]. Following system quality parameters were established.

Table-7: Data of System Suitability Parameter

S.No.	Parameter	Limit	Result
1	Resolution	$R_s > 2$	8.64
2	Asymmetry	$T \leq 2$	Ibrutinib=0.87
3	Theoretical plate	$N > 2000$	Ibrutinib=4689
4	Tailing Factor	$T < 2$	Ibrutinib=1.29

FORCED DEGRADATION STUDIES:

1. Acid Degradation:

A precisely measured 10 mg of unadulterated medication was exchanged to a clean and dry round base cup. 30 ml of 0.1 N HCl was added to it and it was refluxed in a water shower at 60°C for 4 hours. Permitted to cool to room temperature. The sample was then neutralized using dilute NaOH solution & final volume of the sample was made up to 100ml

with water to prepare 100 $\mu\text{g/ml}$ solution. It was injected into the HPLC system against a blank of mobile phase (after optimizing the mobile phase compositions). This experiment was repeated several times using same concentration of HCl (0.1N) and observed its degradation profile. The typical chromatogram shown below is the degradation profile of Minocycline in 0.1N HCl.

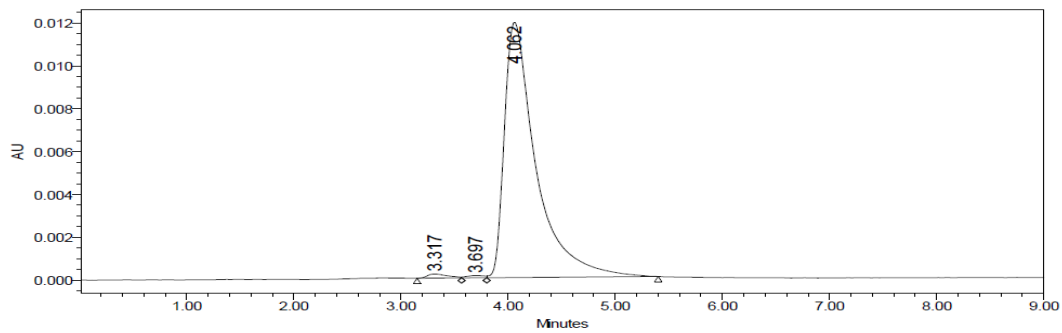


Fig-9: Chromatogram showing degradation for Ibrutinib in 0.1 N HCl

2. Basic Degradation:

An accurately weighed 10 mg of pure drug was transferred to a clean & dry round bottom flask. 30 ml of 0.1N NaOH was added to it. & it was refluxed in a water bath at 60°C for 4 hours. Allowed to cool to room temperature. The sample was than neutralized using 2N HCl solution & final volume of the sample was made up to 100ml to prepare 100

µg/ml solution. It was injected into the HPLC system against a blank of mobile phase after optimizing the mobile phase compositions. This experiment was repeated several times using same concentration of NaOH such as 0.1N to observe its degradation profile. The chromatogram shown below is the degradation profile of Minocycline in 0.1N NaOH.

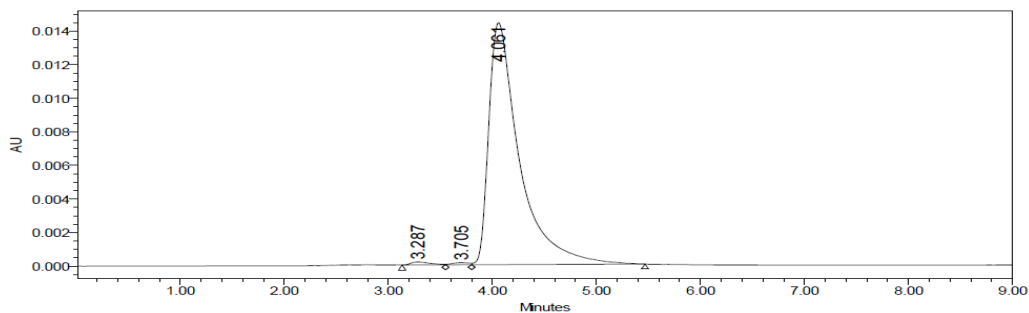


Fig-10: Chromatogram showing degradation related impurity in 0.1 N NaOH

3. Thermal Degradation:

Accurately weighed 10 mg of pure drug was transferred to a clean & dry round bottom flask. 30 ml of HPLC water was added to it. Then, it was refluxed in a water bath at 60° c for 6 hours uninterrupted. After the reflux was over, the drug

became soluble and the mixture of drug & water was allowed to cool to room temperature. Final volume was made up to 100 ml with HPLC water to prepare 100 µg/ml solution. It was injected into the HPLC system against a blank of mobile phase.

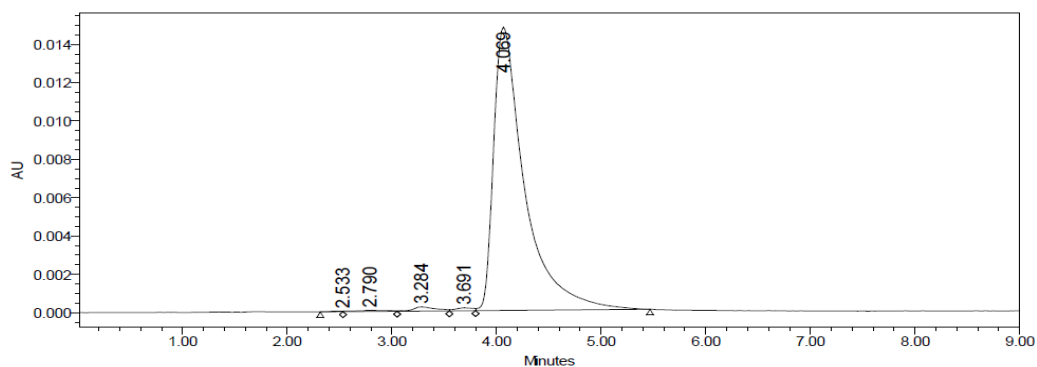


Fig-11: Chromatogram showing thermal degradation studies

4. Photolytic Degradation:

Approximately 10 mg of pure drug was taken in a clean & dry Petri dish. It was kept in a UV cabinet at 254 nm wavelength for 24 hours without interruption. Accurately weighed 1 mg of the UV exposed drug was transferred to a clean & dry 10 ml

volumetric flask. First the UV exposed drug was dissolved in methanol & made up to the mark with mobile phase to get 100 µg/ml solution. Finally, this solution was injected into the HPLC system against a blank of mobile phase and chromatogram was obtained.

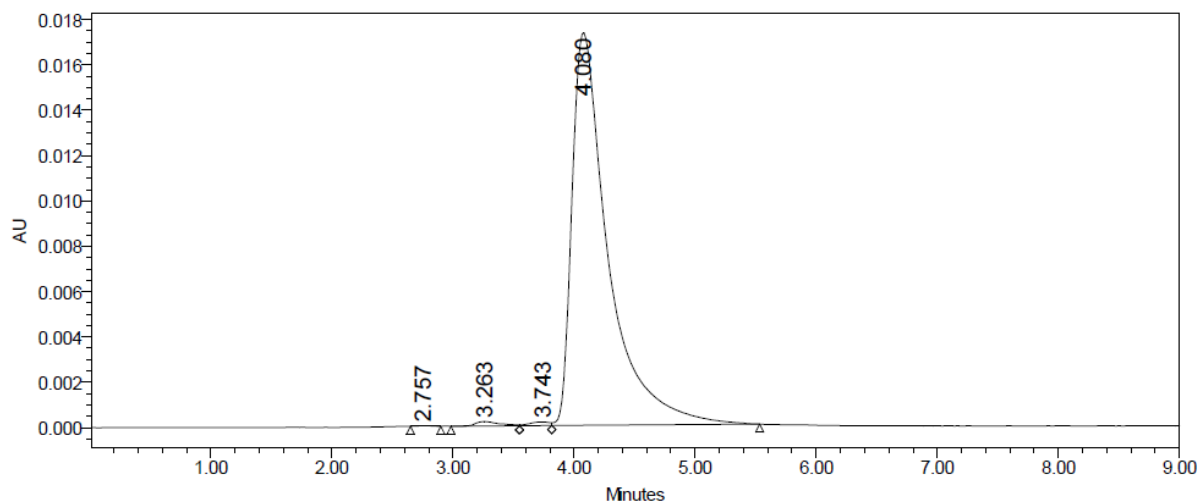


Fig-12: Chromatogram showing photolytic degradation.

5. Oxidation with (3%) H₂O₂:

Accurately weighed 10 mg. of pure drug was taken in a clean & dry 100 ml volumetric flask. 30 ml of 3% H₂O₂ and a little methanol was added to it to make it

soluble & then kept as such in dark for 24 hours. Final volume was made up to 100 ml. using water to prepare 100 µg/ml solution. The above sample was injected into the HPLC system [15].

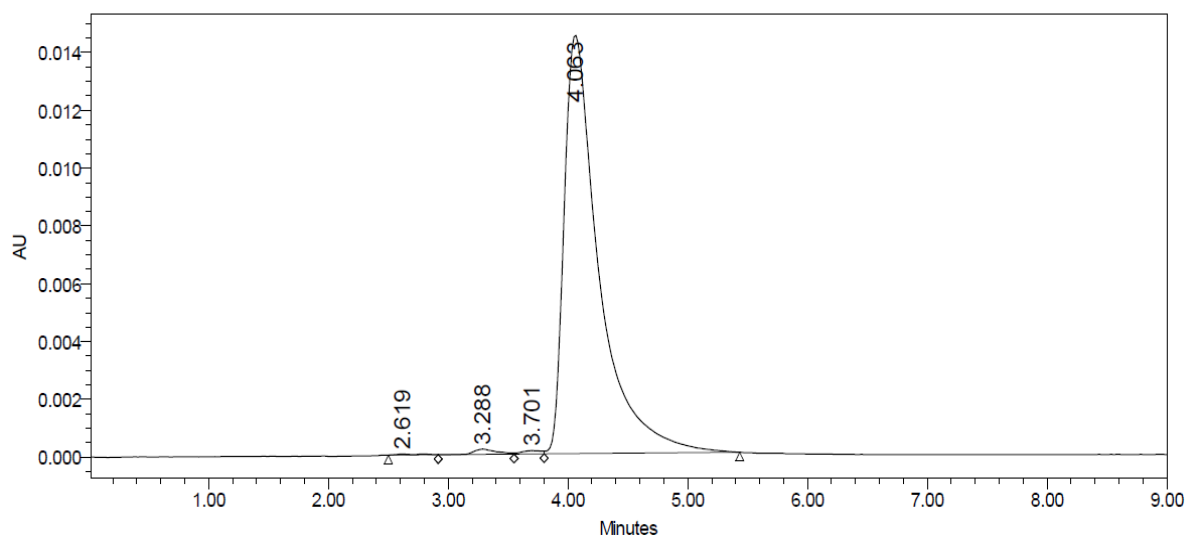


Fig-13: Chromatogram showing oxidative degradation.

Table-8: Results of forced degradation Studies of Ibrutinib.

Stress condition	Time	Assay of substance	of active	Assay of products	of degraded	Mass Balance (%)
Standard	--	100		--		100
Acid Hydrolysis (0.1 M HCl)	24Hrs.	88.32		11.68		100
Basic Hydrolysis (0.1 M NaOH)	24Hrs.	90.04		9.96		100
Thermal Degradation (50 °C)	24Hrs.	91.02		8.98		100
UV (254nm)	24Hrs.	97.25		2.75		100
3 % Hydrogen peroxide	24Hrs.	90.84		9.16		100

CONCLUSION

A sensitive & selective RP-HPLC method has been developed & validated for the analysis of Ibrutinib. Further the proposed RP-HPLC method has excellent sensitivity, precision and reproducibility. The result shows the developed method is yet another suitable method for assay, purity & stability which can help in the analysis of Ibrutinib in different formulations.

REFERENCES:

- Li-min WEI, Zhen-Xing Xu, Peng-fei LV, Yong-le XUE, Xing-Xiang WANG and min ZHANG, A simple HPLC method for the determination of Ibrutinib in rabbit plasma and its application to a pharmacokinetic study. *Latin American journal of pharmacy*, 2016; 5(1): 130-34.
- Pan Z, Scheerens H, Li SJ, Schultz BE, Sprengeler PA, Burrill LC, Mendonca RV, Sweeney MD, Scott KC, Grothaus PG, Jeffery DA, Spoerke JM, Honigberg LA, Young PR, Dalrymple SA, Palmer JT (Jan 2007). "Discovery of selective irreversible inhibitors for Bruton's tyrosine kinase". *ChemMedChem* 2(1): 58-61.
- Jump up^ US patent 7514444, Honigberg L, Verner E, Pan Z, "Inhibitors of Bruton's Tyrosine Kinase", *pharmacyclics Inc* 7 April 2009, issued 28 December 2006.
- Morgan, David J. "Fraction collector (post on Flickr)". Flickr. Retrieved 28 October 2015.
- Karger, Barry L. (1997). "HPLC: Early and Recent Perspectives". *Journal of Chemical Education*. 74: 45. Bibcode:1997JChEd..74...45K.
- Henry, Richard A. (1 February 2009) "The Early Days of HPLC at Dupont". *Chromatography Online*. Avanstar Communications Inc.
- Iler, R.K. (1979) *The Chemistry of Silica*. John Wiley & Sons. New York.
- Karger, B. L.; Berry, L. V. (1971). "Rapid liquid-chromatographic separation of steroids on columns heavily loaded with stationary phase". *Clin. Chem.* 17 (8): 757-64.
- Giddings, J. Calvin (1965) *Dynamics of Chromatography, Part I. Principles and Theory*. Marcel Dekker, Inc., New York. p. 281.
- Ettre, C. (2001). "Milestones in Chromatography: The Birth of Partition Chromatography" (PDF). *LCGC*. 19 (5): 506-512. Retrieved 2016-02-26.
- Martin, A J P; Synge, R L M (1941). "Separation of the higher monoamino-acids by counter-current liquid-liquid extraction: the amino-acid composition of wool". *Biochemical Journal*. 35 (1-2): 91-121.
- Lindsay, S.; Kealey, D. (1987). *High performance liquid chromatography*. Wiley. from review Hung, L. B.; Parcher, J. F.; Shores, J. C.; Ward, E. H. (1988). "Theoretical and experimental foundation for surface-coverage programming in gas-solid chromatography with an adsorbable carrier gas". *J. Am. Chem. Soc.* 110(11): 1090-1096.
- Displacement Chromatography. *Sacheminc.com*. Retrieved 2011-06-07. Archived September 15, 2008, at the Wayback Machine.
- Snyder, Lloyd R.; Dolan, John W. (2006). *High-Performance Gradient Elution: The Practical Application of the Linear-Solvent-Strength Model*. Wiley Interscience. ISBN 0470055510.
- Majors, Ronald E. (2010-09-07) *Fast and Ultrafast HPLC on sub-2 µm Porous Particles — Where Do We Go from Here? — LC-GC Europe*. *Lcgeurope.com*. Retrieved 2011-06-07.