



PHARMACOLOGICAL EVALUATION OF POLYHERBAL EXTRACTS OF *Amorphophallus paeoniifolius*, *Citrus sinensis* AND *Plumbago zeylantia* FOR ANTI FUNGAL ACTIVITY

Matangi Suryaprabha* and Nadendla Rama Rao

Department of Biotechnology, Chalapathi Institute of Pharmaceutical Sciences, Lam, Guntur-522034

*Corresponding Author Email: surya_pharm@yahoo.com

ABSTRACT

Fungal infections may take the form of superficial infections involving the skin or mucous membranes or systemic involving various internal organs. Deep infections are systemically treated with drug therapy. Among the fungi that cause systemic infections are *Aspergillus* spp., *Nocardia asteroides*, *Histoplasma capsulatum* and *Coccidioides immitis*. Among ancient civilizations, India has been known to be rich repository of medicinal plants. Recently, WHO estimated that 80 percent of people worldwide rely on herbal medicines for some aspect of their primary health care needs. Treatment with medicinal plants is considered very safe and there is no or minimal side effects. Medicinal plants are considered as rich sources of ingredients which can be used in drug development. The Ayurvedic literature *Sarangdhar Samhita* highlighted the concept of polyherbalism to achieve greater therapeutic efficacy. The active phytochemical constituents of individual plants are insufficient to achieve the desirable therapeutic effects. When combining the multiple herbs in a particular ratio, it will give a better therapeutic effect and reduce the toxicity. This research mainly focuses on important of the polyherbalism and its clinical significance.

KEY WORDS

Polyherbals, Fungal infections, *Amorphophallus Paeoniifolius*, *citrus sinensis*, *Plumbago Zeylantia*.

INTRODUCTION

Infectious diseases represent a critical problem to health, and they are one of the main causes of morbidity and mortality worldwide (World Health Organization 1998). Despite the significant progress in human medicine, infectious diseases caused by microorganisms such as fungi are still a major threat to public health. The impact is even more in developing countries due to the unavailability of medicine and the emergence of widespread drug resistance.

One of the major public health problems is Dermatophytosis. Dermatophytes include three types of fungus such as *Microsporum*, *Trichophyton* and *Epidermophyton* which commonly causes skin diseases in animals and humans. Dermatophytosis is an

infectious disease of skin, hair and nails which attack on the keratinized tissue.

The rich content of antifungal substances in plants are being used biopesticide since up to the beginning of human civilization. Antifungal effects of plant and plant products emerge clearly every day. Antifungal substances which are obtained from plants have no side effect against environment thus, giving a significant advantage. There are many synthetic and natural product-based drugs available for treating fungal infections, but they are not consistently effective. Furthermore, the development of resistance in fungi against most of the drugs has now been reported for several years. The use of amphotericin B, known as the "gold standard", is limited because of its infusion-related problems and nephrotoxicity. In addition, the low efficacy, and side-effects and resistance associated

with the existing drugs, highlight the advent of safe, novel, and effective antifungal drugs. Plants produce a great deal of secondary metabolites, many of them with antifungal activity. Similarly, traditional medicine has made use of many different plant extracts for treatment of fungal infection and many of these have been tested for *in vitro* antifungal activity. Based on the knowledge that plants develop their own defense against fungal pathogens, they appear as an interesting source for antifungal compounds.

MATERIAL AND METHODS

Amorphophallus

paeoniifolius (Dennst.) Nicolson (Araceae) is a commonly available tuber in South India, widely used in folk medicine for treatment of acute rheumatism, tumors, lung swelling, asthma, vomiting, and abdominal pain. So far, no attempts have been made to evaluate the chemical composition and medicinal properties of *A. paeoniifolius*. Hence, the present study was performed to investigate the antifungal potential of ethanol extract of *A. paeoniifolius* using different *in vitro* models.

Citrus sinensis L. is an evergreen tree belonging to the family Rutaceae family. It is native to China but is widely cultivated throughout the world. It currently ranks as the most commonly cultivated and commercialized specie of citrus. Citrus fruits, including their by-products, have been reported to possess high medicinal value in addition to their economic values. The fresh fruits are consumed on a large scale and large quantities are processed to produce juice. The wastes produced, after consumption and juice extraction, such as the peels, pulps and seeds are a potential source of valuable byproducts. The oil obtained from the seeds, flowers, fruits and rinds of different species of Citrus also find wide applications in the toiletry, confectionary, and perfumery industry. The oils extracted from the peels and seeds of *C. sinensis* have also been reported to possess different type of activities ranging from insecticidal activity, to antimicrobial activity against a wide range of microbial organisms, hence this study was designed to evaluate and validate the activity of *C. sinensis* seed oil through an *in vitro* antifungal assay.

Plumbago zeylanica L. is a multipurpose medicinal herb of family Plumbaginaceae. A native of South Asia, the species is distributed throughout most of the tropics

and subtropics. In India *P. zeylanica* commands an important place among medicinal herbs in India since ancient times. Ayurveda, the Indian indigenous system of medicine dating back to the Vedic ages (1500-8000 BC), has described chitraka as tumor-negating and anti-dyspeptic. Plumbagin has shown antibacterial activity against both gram-positive (e.g. *Staphylococcus*, *Streptococcus*, *Pneumococcus* sp.) and gram-negative (e.g. *Salmonella*, *Neisseria*) bacteria. It is also active against certain yeasts and fungi (*Candida*, *Trichophyton*, *Epidermophyton* and *Microsporum* spp.) and protozoa (*Leishmania*).

Preparation of extracts

Air dried coarsely powdered plant materials of (*Amorphophallus Paeoniifolius*, *Citrus Sinensis* and *Plumbago Zeylantia*) were extracted with ethanol (95%) using soxhlet apparatus for 4-5 hrs. All the extracts were concentrated at low pressure by rotary flash evaporator and finally air-dried.

Microbial cultures and Growth Stains

The plant extracts were assayed for antifungal activity against the fungal strain *Bacillus subtilis* (Gram +ve) and *Escherichia coli* (Gram -ve).

Potato Dextrose Agar (PDA) Medium (pH 6.7)

Potato - 250g

Dextrose - 15g

Agar - 18g

Distilled water - 1000ml

Effect of Plant Extracts on Mycelium Growth

The poisoned food method was used in the preliminary screening of aqueous extracts for their antifungal properties evaluation. First, the mycelia growths were evaluated in 60 mm Petri dishes filled with PDA solid medium amended with 10% and 20% aqueous extracts of each plant. Next, the center of each Petri dish was inoculated with 5 mm diameter disc of fungal mycelium, taken from pure culture (7 days old). Then, all inoculated dishes were incubated at 25° C for 6 days. After that, the radial mycelial growth was measured 6 days after inoculation. For each treatment, three replicates were maintained.

Finally, the antifungal activity of each extract was calculated in terms of inhibition percentage of mycelia growth by using the following formula:

$$\% \text{ inhibition} = (dc - dt) / dc \times 100$$

where *dc* is the average increase in mycelia growth in control and *dt* is the average increase in mycelia growth in treated.

RESULTS

Table 1: Effects of 10% and 20% concentrations of aqueous extracts of different plants on growth of *Asperigillus* and *C albicans* for mycelia growth index.

Treatment	Concentration	Mycelia growth index (mm/hr)		Mean
		<i>Asperigillus</i>	<i>C albicans</i>	
Control	0%	0.64 ± 0.015	0.70 ± 0.028	0.67
<i>Amorphophallus Paeoniifolius</i>	10%	0.58 ± 0.027	0.61 ± 0.012	0.60
<i>Amorphophallus Paeoniifolius</i>	20%	0.51 ± 0.052	0.47 ± 0.024	0.54
<i>Citrus Sinensis</i>	10%	0.60 ± 0.041	0.63 ± 0.054	0.61
<i>Citrus Sinensis</i>	20%	0.56 ± 0.072	0.61 ± 0.038	0.58
<i>Plumbago Zeylantia</i>	10%	0.62 ± 0.036	0.68 ± 0.018	0.65
<i>Plumbago Zeylantia</i>	20%	0.57 ± 0.023	0.62 ± 0.056	0.59
Polyherbal Extract	10%	0.43 ± 0.072	0.54 ± 0.012	0.48
Polyherbal Extract	20%	0.41 ± 0.052	0.53 ± 0.024	0.47

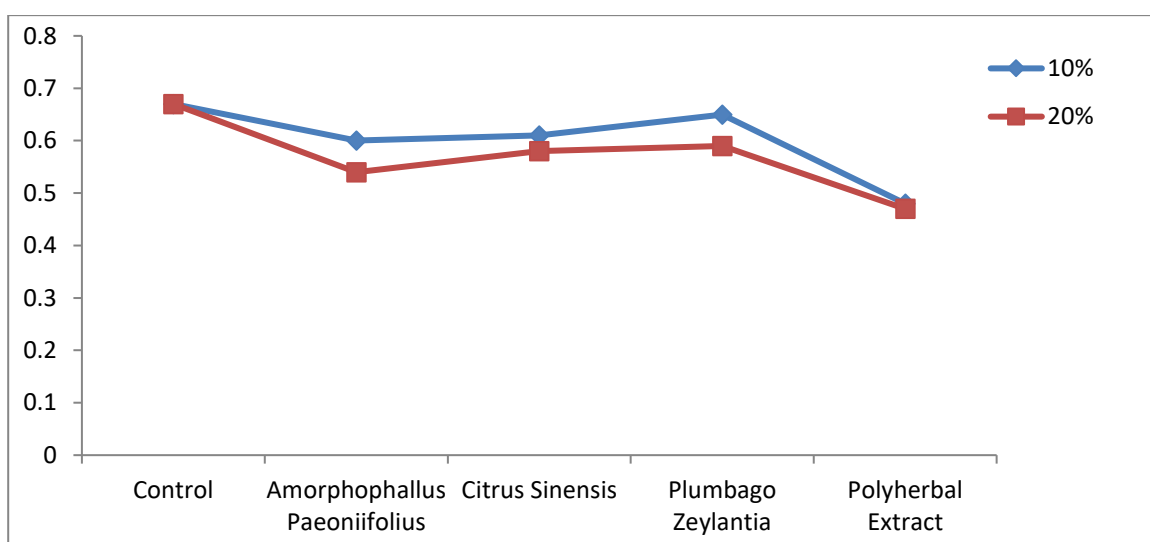


Figure 1: Mycelia growth index of *Asperigillus* and *C albicans* on various treatments

DISCUSSION

This study showed the treatment with various extracts on *Asperigillus* and *C albicans* results in anti-fungal activity when compared with control. The extracts of *Amorphophallus Paeoniifolius*, *Citrus Sinensis* and *Plumbago Zeylantia* showed anti-fungal activity with very much little variance and when the combination of extract (polyherbal extract) was given, results in gradual enhancement in anti-fungal activity. This shows the synergistic activity achieved by polyherbal extracts.

CONCLUSION

The Polyherbal Extracts of *Amorphophallus Paeoniifolius*, *Citrus Sinensis* and *Plumbago Zeylantia* for Anti-Fungal Activity was evaluated and was found to be very potent regarding anti-fungal activity when they were given in combination.

REFERENCES

1. S. Satish, D. C. Mohana, M. P. Ranhavendra, and K. A. Raveesha, "Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* sp.," *Journal of Agricultural Technology*, vol. 3, no. 1, pp. 109–119, 2007.
2. Nakamura CV, Ishida K, Faccin LC, Filho BP, Cortez DA, Rozental S, et al. *In vitro* activity of essential oil from *Ocimum gratissimum* L. Against four *Candida* species. *Res Microbiol* 2004; 155:579-86.
3. Naglik JR, Challacombe SJ, Hube B. *Candida albicans* secreted aspartyl proteinases in virulence and pathogenesis. *Microbiol Mol Biol Rev* 2003; 67:400-28.
4. Jenie BS. Antimicrobial Activity of *Piper betle* Linn Extract Towards Foodborne Pathogens and Food Spoilage Microorganisms, FT Annual Meeting, New Orleans, Louisiana; 2001.

5. Clinical and Laboratory Standards Institute. CLSI Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Approved Standard. CLSI document M27-A3. 3rd ed. Wayne: Clinical and Laboratory Standards Institute; 2008a
6. P. V. Nielsen and R. Rios, "Inhibition of fungal growth on bread by volatile components from spices and herbs, and the possible application in active packaging, with special emphasis on mustard essential oil," *International Journal of Food Microbiology*, vol. 60, no. 2-3, pp. 219–229, 2000.
7. D. Bhatnagar and S. P. McCormick, "The inhibitory effect of neem (*Azadirachta indica*) leaf extracts on aflatoxin synthesis in *Aspergillus parasiticus*," *Journal of the American Oil Chemists' Society*, vol. 65, no. 7, pp. 1166–1168, 1988
8. V. Terzi, C. Morcia, P. Faccioli, G. Vale, G. Tacconi, and M. ` Malnati, "In vitro antifungal activity of the tea tree (*Melaleuca alternifolia*) essential oil and its major components against plant pathogens," *Letters in Applied Microbiology*, vol. 44, no. 6, pp. 613–618, 2007
9. A. Ergene, P. Guler, S. Tan, S. Mirici, E. Hamzaoglu, and A. Duran, "Antibacterial and antifungal activity of *Heracleum sphondylium* subsp. *Artvinense*," *African Journal of Biomedical Research*, vol. 5, no. 11, pp. 1087–1089, 2006.
10. D. C. Mohana, K. A. Raveesha, and K. M. L. Rai, "Herbal remedies for the management of seed-borne fungal pathogens by an edible plant *Decalepis hamiltonii* (Wight & Arn)," *Archives of Phytopathology and Plant Protection*, vol. 41, no. 1, pp. 38–49, 2008.

Received:04.08.18, Accepted: 07.09.18, Published:01.10.2018

***Corresponding Author:**

Matangi Suryaprabha*

Email: surya_pharm@yahoo.com