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Evaluation of Anti-Inflammatory Activity of Aerial Part Extract of Nerium Indicum in Acute and Chronic in vivo Models in Mice

- ^{1*}Nirmala Devi, ¹Sunil Kumar Prajapati and ²Ajay Kumar Gupta
- *1PhD Research scholar at School of Pharmacy, Monad University, Hapur, UP, INDIA
- ¹Institute of Pharmacy, Bundelkhand University, UP, India.
- ²University Institute of Pharmacy, CSJM University, UP, INDIA.

Received: 10 Oct 2018 / Accepted: 8 Nov 2018 / Published online: 1 Jan 2019 Corresponding Author Email: nirmalaprajapati05@gmail.com

Abstract

Medicines from natural sources have become a mainstay in drug discovery. Biocompatibility and lower side effects of natural drugs from plants, animals, microbes and their derivatives, are the main horizons to screen such drugs. In this study aerial parts of Nerium indicum were extracted together in ethanol and distilled water. Both extract were screened for anti-inflammatory potential using three models viz., Carrageenan induced Paw edema, Cotton pellet induced Granuloma and Carrageenan induced Peritonitis. The maximum inhibition was observed for at the dose of 400 mg/kg (49.39%) and minimum inhibition at the dose of 100mg/kg (1.25%) after 4h of drug treatment in Carrageenan induced paw edema, whereas diclofenac sodium (standard drug) produced 59.03% inhibition. In the chronic model (cotton pellet induced granuloma), ethanolic extract of N.indicum (at doses of 100, 200 and 400 mg/Kg), Diclofenac as standard drug showed decreased formation of granuloma tissue by 32.84%, 39.18%, 42.07% and 67.98% respectively. In addition, exudative model of inflammation was mimicked using carrageenan induced Peritonitis. Percentage of Leukocyte and Neutrophil inhibition was calculated. Dose related inhibition was observed. At 400 mg/kg N.indicum inhibited 42.69 % leukocytes and 44.05 % Neutrophils. Thus, the results indicate that ethanolic extract of N.indicum aerial parts have potent anti-inflammatory effects on both acute and chronic type of inflammation in mice.

Keywords

N.indicum, Carrageenan induced paw edema, Peritonitis, Cotton Pellet granuloma.

INTRODUCTION

Plants are among the most widely sought source of therapeutics. The battle of existence is similar for plants and any other form life on earth. Mankind has always been curious about unearthing the mysteries behind this similarity. Exploring and exploiting the essence of healthy living from nature has achieved many milestones till date. Advanced understanding of air, water, earth, matter and life has enriched the ways of development and discovery. Every range of product for health and wellbeing constitute natural elements for e.g. morphine, quinine, colchicine, atropine, pilocarpine, or theophylline. Paclitaxel, Vinca alkaloids, theophylline, Colchicine, capsaicin curcumin, resveratrol, epigallocatechin-3-gallate (EGCG), and quercetin are other important plant



products. Examples for plant-derived compounds that served as lead structures and/or were chemically modified are salicylic acid (acetylsalicylic acid), morphine (scores of derivatives), camptothecin (topotecan and irinotecan), artemisinin (artemether), and dicoumarol (warfarin) [1]

Supplementation of core therapeutics with Adjuvant is improving the survival ratio of people with serious debilitating diseases like cancer, leprosy, Aids, flu, arthritis diabetes and infections etc. a huge list of medicinal plant and their products are in market, under clinical and preclinical trials [2]. Thorough congregation of information, knowledge and skills from various literary compilations, folkloric and traditional practitioners and internet tools has led the development of a new trend known as 'Reverse Pharmacology' [3].

Inflammation is a sign of active defense machinery. Sometimes a cascade of inflammatory mediators leads to damage and abrupt changes in the body [4]. Conventional anti-inflammatory drugs include NSAIDs, corticosteroids, immunosuppressant and other biological. All of these produce serious and numerous toxicities [5]. Demand for inflammatory agents depends on the source of inflammation and this generates the interest among scientific community to search for newer agents. Likewise, antimicrobials are always an important research due to the increased rate of resistance to existing antimicrobials. Discovery and development of safe and effective antimicrobials carry an immense therapeutic relevance.

Transcription factor $NF\kappa B$ considered for anti-inflammatory activity.

Acute and chronic inflammations are induced by chemical mediators such as prostaglandins, leukotrienes and platelet-activating factor. Antiinflammatory agents show their activities through several activity mechanisms [6]. Non-steroid antiinflammatory drugs (NSAIDs) are the most prescribed drugs for treatment of inflammatory diseases. The NSAIDs provide the patients with symptomatic relief; however, they do not modify the pathogenesis of inflammation [7]. Furthermore, prolonged use should be avoided due to severe side effects particularly on gastric mucosa. Therefore, searching for new drug candidates in the treatment of chronic inflammation has great importance. The scientific research on the biological effects of the medicinal plants led to the discovery and development of novel bioactive drug molecules. Indeed, natural products have proved to be a rich source of therapeutic agents. Due to the side effects

caused mostly by synthetic drugs, research into natural products has advanced tremendously in academia and pharmaceutical companies. The consumer interest in such plant-based remedies is due to the lower cost of phytotherapy and the fact that many plant-based remedies are successfully replacing allopathic medicines in relieving malady symptoms.

For centuries, in rural areas, the efficacious plants have been used in simple formulations to treat several diseases. Inflammatory diseases are among the most common health problems treated with traditional remedies. In vivo and in vitro antiinflammatory, antinociceptive and antipyretic evaluations of medicinal plants provide scientific evidence for the ethnomedicinal features. In the past few years, many studies have investigated the potential of healing higher plants ethnobotanical histories. Many such plant-derived molecules have been isolated, identified and successfully introduced into international markets by pharmaceutical industries [8]. Particularly phenolic plant constituents such as flavonoids, phenolic acids and proanthocyanins were reported to modulate arachidonic acid metabolism. Similarly, a number of phenolic compounds were shown to be the active anti-inflammatory and antinociceptive compounds of folk remedies through bioassay-guided isolation procedures [9].

In an in vitro study, inhibitory activity against PGE1and PGE2-induced contractions [10].Antiinflammatory and antinociceptive activities of isolated compounds [11].

Importantly, the anti-inflammatory potency of secondary metabolites like, flavonoids was found to be [12], Alkaloids [13], Phenolics [14], Tannins, Glycosides [15]. In vitro inhibitory activity against both thromboxane and leukotriene synthesis [16]. In-vitro inhibitory activity against tumor necrosis factor α (TNF- α) production [17].

It is obvious that folk medicines are the most favorable sources for novel drug discovery. By using in vivo bioassay- or in vitro activity-guided fractionation and isolation techniques, hundreds of active constituents have been identified. Hence, it would be reasonable to follow an ethnopharmacological approach using traditional knowledge to increase success in drug discovery and development.

If unchecked, inflammation leads to development of rheumatoid arthritis, diabetes, cancer, Alzheimer's disease, and atherosclerosis along with pulmonary, autoimmune and cardiovascular diseases. Inflammation involves a complex network of many



mediators, a variety of cells, and execution of multiple pathways. e activated immune system, including activated immune cells and the biomolecules. Furthermore, chronic inflammation is also linked with various steps of tumorigenesis and recognized as risk factor for the occurrence of different types of cancers [18]. The chronic use of is connected with cardiovascular, gastrointestinal, and renal toxicities [19-20]. Similarly, the use of corticosteroids leads to hypertension, hyperglycemia, osteoporosis, and growth arrest [21]. Natural products are safe, efficacious, biocompatible, and cost-effective alternatives to treat inflammatory diseases [22]. The anti-inflammatory effects of phytoconstituents are exerted through their action on key regulatory molecules, including cyclooxygenase (COX), inducible nitric oxide synthase (iNOS), and cytokines.

Nerium indicum is one of the Apocynaceae family plants. There are varieties of Nerium in India alone. It is known for its traditional and conventional therapy of [23-28]. The plant is rich in toxic secondary metabolites which is a characteristic of Apocynacaea family [my all]. Presence of cardiac glycosides, alkaloids, terpenoids, antioxidants, tannins etc [29-30]. Has made the plant very sourceful to treat the aforementioned ailments. The main chemical constituents are, Anvirzel, noeflx a, olendrin, odoroside, are used in the [31-32].

"To date the best source of anticancer agents have come from toxic plants". Based on traditional claims, Arrow poisons are anticancer [33].

This research article reports Invitro antioxidant, antimicrobial, antiproliferative and antiinflammatory activity in mice. Reporting above activities on the aerial parts extract of pink variety of Nerium indicum Mill found in Jhansi region of Bundelkhand is not done previously.

MATERIAL AND METHODS Collection and authentication of plant

The plant growing wildly at Jhansi region of Bundelkhand was collected in the month of April. The Plant were identified and authenticated by Dr. Mudailiya, taxonomist NVARI, Jhansi, UP India. Herbarium specimen no. 24381 for future reference.

Plant extract preparation

The parts of plants (leaves, flowers, fruit follicles and stem) washed thoroughly and dried. Pulverized and sieved drug was extracted exhaustively in Soxhlet apparatus using water, 60% ethanol and ethanol as solvent. The extracts were concentrated using vacuum rotary evaporator and Lyophilizer and dried

in incubator. Percentage yield calculated and stored in deep freezer (-4°C) for future use.

Phytochemical study

Qualitative analysis of Phytoconstituents performed according to standard methods [34-35].

Antiinflammatory Study

Animals

Animal study protocol was approved by the Institutional Animal Ethics Committee (IAEC), Bundelkhand University [Project BU/Pharm/IAEC/a/16/15]. Adult healthy female and male Swiss Albino mice, of age and body weight (6-8 week, 25-35g) were used in the study. Animals were housed in the Institutional Animal house facility. Guidelines of CPCSEA (Regd. No. 716/02/a/CPCSEA) and IAEC were strictly followed for the care and use of animals. All animals were provided sanitized paddy husk bedding and fed with nutrient rich diet and water ad libitum. Mice were kept in polypropylene lab cages in a room conditioned with a12h light: dark cycle, temperature at 20 ± 2°C and relative humidity 54% \pm 12%. The animals were tamed and caressed for one or two weeks before use. Acute toxicity Study [36-38]:

The limit test for acute toxicity was carried out at 2000 mg/kg oral dose of Nerium extract as per OECD 423 guidelines (OECD 423/425, 2003). Swiss albino female mice (20-30g) were fasted for 6 hrs prior to dosing. Each single dose (1ml) was administered in three mice by oral gavage feeding needles for mice. The mice were observed for any change in the behavior, neurological profile and autonomic profile keenly watched at 24, 48 and 72 hrs for any comorbidity or morbidty.

Drugs/chemicals

Diclofenac, carrageenan, and dextran were purchased from Sigma Aldrich (USA) and formalin was purchased from CDH Mumbai. All other reagents were of analytical grade

Paw edema induced by Carrageenan [39-40]

Carrageenan-induced acute inflammation in the mice paw is used as a classical model of edema formation and hyperalgesia, for the study of non-steroidal anti-inflammatory drugs and selective cyclooxygenase-1 (COX-1) and COX-2 inhibitors.

Five groups of 6 mice each were treated with Diclofenac sodium (10 mg/kg, p.o), or vehicle (distilled water, p.o). One hour after receiving the drug(s), each animal received a subcutaneous injection of 0.02 ml of 1%w/v λ -carrageenan (dissolved in 0.9% NaCl) in the right hind paw of each mouse under the subplantar aponeurosis. The paw volume was measured by dipping the foot in the mercury bath of a plethysmometer up to the



anatomical hairline on lateral malleolus and compared with control animals at 0, 1, 2, 3 and 4 h interval. The difference between the final and initial

volumes used to calculate Percent inhibition according to the following formula:

Percentage inhibition = (Ct- C₀)_{control} - (Ct- C₀)_{treated} / (Ct- C₀)_{control} × 100

Where, Ct=mean paw volume for each group at time t, and C₀=mean paw volume for each group before carrageenan injection.

Cotton Pellet Induced Granuloma [41-42]

This study was carried out following the protocol of Ray SD, 2015 and Ismail TS, 1997 [26] with fewer modification. Sterile cotton pellets (10 ± 0.5 mg) were implanted subcutaneously on the dorsa region of mice. The five groups (n = 6/group) of mice were treated with Normal saline, Extract (100, 200 mg/kg and 400 mg/kg) and Diclofenac (10 mg/kg) orally, once daily over 7 consecutive days. On day 8, the mice were anaesthetized, and the implanted cotton pellets were taken out carefully. The pellets coated with granulomatous tissue were dried overnight at 60 °C and weighed. The mean difference between the weight of control and treated animal in each group was calculated. And precent inhibition calculated as above.

Peritonitis induced by carrageenan [43-44]

This model predicts neutrophil migration to the peritoneal cavity and leukocyte rolling. Mice were treated Diclofenac (10 mg/kg, po) or vehicle (distilled water, po). One hour after treatment, each animal received an intraperitoneal injection of 0.25 mL 1% carrageenan to induce the inflammatory process and, 4 h later, the animals were sacrificed by cervical dislocation after CO₂ euthanasia. The peritoneal cavity was washed with 2 mL phosphate-buffered

saline (PBS) containing heparin, and the exudates/peritoneal cavity cells were collected for analysis. The volume recovered was similar in all experimental groups and equated to approximately 95% of the injected volume. An exudates sample of 20 μ L was taken, and a 0.4-mL Turk's solution was added to it. The total number of neutrophils per cavity was then counted in a Neubauer chamber using light microscope. Data are reported as the mean (x $10^3/\text{mm}^3$) number of leukocytes and neutrophils.

Statistical analysis of Data

Statistical Calculations were performed using GraphPad Instat 3.0. Data are reported as means ± SEM for 6 animals per group. Statistical analyses were carried out using one-way analysis of variance (ANOVA), followed by the Student-Newman-Keuls post hoc test for multiple comparisons. P values < 0.05 were considered to be statistically significant.

RESULT AND DISCUSSION

The results obtained are provided in Table 1-4 and Graphs 1-3. Phytocontituents analysis (Table 1) showed the presence alkaloids, glycosides, terpenoids, phytosterols, tannins etc.

Table 1: Preliminary Phytoconstituents analysis

S.NO.	Test	Nerium indicum		
Tests	Extracts	N1	N2	N3
Glycosides-cardiac/anthraquinone	Borntrager's test	++	++	+/-
Saponins	Foam test	-	-	++
Oils and fat	Spot test	+++	++	+
Phlobatannins/ Chalcones	HCl test/spot test	-	-	-
Flavonoids	AlCl₃ test/ alkaline reagent test/lead acetate test	+++	+++	+++
Tannins/Phenolic compound	FeCl₃ test/ Lead acetate test	+++	+++	+++
Alkaloids	wagner's test/mayer's test/hagers/dragendorffs	++++	++++	++++
Protein/Amino acids	ninhydrin/ xanthoproteic	++	++	++
Steroids	salkowski's test/ Liebermann buchard	-	-	-
Phytosterols	sulphuric acid test	+	-	-
Carbohydrates/ sugar	molish's test/ Benedict's test	+	-	+
Coumarins	fluorescens test	+	-	-



Table 2: Carrageenan induced rat paw oedema: (n=6)

		Percentage inhibition							
iroup/ eatme Dose		1 hour		2 hours		3 hours		4 hours	
Group/ Treatment	۵	Volume (ml)	% Inhibition						
Group-I (Control)	5 ml/ kg	0.87±0.03		0.89±0.04		0.84±0.03		0.83±0.03	
Group-II (Diclofenac)	10 mg/kg	0.36±0.03***	58.62%	0.39±0.03***	56.17%	0.32±0.02***	61.90%	0.34±0.01***	59.03%
Group-III	100 mg/kg	0.83±0.05 ^{\$\$\$}	4.58%	0.85±0.05 ^{\$\$\$}	4.49%	0.80±0.05 ^{\$\$\$}	4.76%	0.82±0.05 ^{\$\$\$}	01.25%
Group-IV	200 mg/kg	0.83±0.06 ^{\$\$\$}	4.58%	0.84±0.05 ^{\$\$\$}	5.61%	0.81±0.06 ^{\$\$\$}	3.75%	0.79±0.07 ^{\$\$\$}	4.81%
Group-V	400 mg/kg	0.45±0.07***\$\$	48.27%	0.46±0.07***\$\$\$	48.31%	0.43±0.08***\$\$\$	48.80%	0.42±0.06***\$\$\$	49.39%

The percent inhibition for each group was calculated by comparison with the control group. Values indicate mean±S.E.M (ANOVA test followed by Tukey's test). *** Data differs significantly (p≤0.0001) when compared against control group.

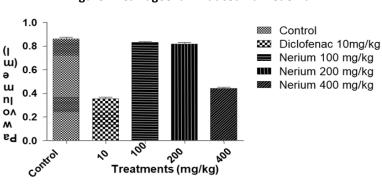
Table 3: Cotton pellet induced Granuloma: (n=6)

Group/ Treatment	Dose	Wt of cotton Mean ±S.E.M.	% Protection	
Group-I Control (Saline)	5ml/kg	62.20±6.10		
Group-II (Diclofenac)	10mg/kg	19.93±2.05***	67.98%	
Group-III	100mg/kg	41.77±3.75 ^{\$\$\$}	32.84%	
Group-IV	200mg/kg	37.83±2.1 ^{\$\$\$}	39.18%	
Group-V	400mg/kg	36.03±2.25*** ^{\$\$\$}	42.07%	

*** Data differs significantly (p≤0.0001) when compared against control group Table 3: Carrageenan Induced Peritonitis: (n=6)

	Dose	Parameters				
Group/ Treatment		Leucocytes (10 ⁵ mm ⁻³)	Leucocytes inhibition	Neutrophils (10 ⁵ mm ⁻³)	Neutrophils inhibition	
Group-I Control (Saline)	5ml/kg	4.45±0.35		2.02±0.03		
Group-II (Diclofenac)	10mg/kg	2.02±0.12***	54.60%	0.92±0.02***	54.45%	
Group-III	100mg/kg	3.96±0.31 ^{\$\$\$}	11.01%	1.94±0.04 ^{\$\$\$}	3.96%	
Group-IV	200mg/kg	3.85±0.25 ^{\$\$\$}	13.48%	1.95±0.02 ^{\$\$\$}	3.46%	
Group-V	400mg/kg	2.55±0.19***\$\$\$	42.69%	1.13±0.02*** ^{\$\$\$}	44.05%	

^{***} Data differs significantly (p≤0.0001) when compared against control group



Treatments (mg/kg)

Figure 1: Carrageenan induced Paw edema





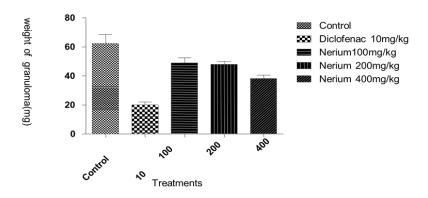
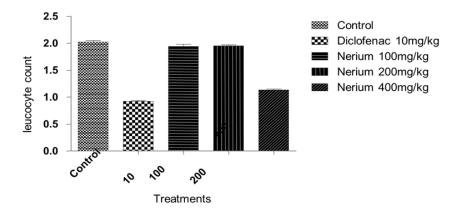


Figure 3: Carrageenan induced peritonitis



The rationale behind this work was to scientifically authentify and confirm the traditional, folk and preliminary claims of Nerium indicum for its antiinflammatory activity. Results of the present study indicate that the extract of N.indicum posses significant anti-inflammatory activity when used in doses of 100, 200 and 400 mg/kg, both in acute and chronic models of inflammation. The significant activity observed in the suppression of carrageenan induced acute model of inflammation extract as well as by diclofenac, may be due to inhibition of release of various mediators released during different phases of carrageenan induced inflammation [45]. Development of oedema in the rat paw after injection of carrageenan is a biphasic event. The early phase (1-2 h) is mainly mediated by histamine, serotonin and maintained during the plateau phase by kinin and kinin-like substances. The second accelerating phase of swelling is due to the increased synthesis and release of prostaglandins, protease and lysosome in damaged tissue surrounding [46]. Paw swelling, or footpad edema, is a convenient method for assessing inflammatory responses to

antigenic challenges and irritants eg carrageenan, croton oil, Sheep Red Blood Cells etc. This model has long been used to assess the anti-inflammatory properties of agents such as nonsteroidal anti-inflammatory drugs (NSAIDs) that inhibit prostaglandin production.

The cotton pellet granuloma method has been widely employed to assess the transudative, exudative and proliferative component of chronic inflammation. It can be noted from the foreign body induced granuloma studies that the extract at the dose of 400 mg/kg showed 42.07% inhibition whereas diclofenac showed 67.98% inhibition in the mean dried weight of granuloma. These results also indicate that the extract possess dose dependent anti-transudative activity.

Nerium reduces the neutrophil migration in carrageenan-induced peritonitis in mice. Mice were pretreated subcutaneously (1 min) with saline (Control), ethanolic extract of N.indicum (100, 200 and 400 mg/kg) and received an intraperitoneal injection of carrageenan (0.2 ml/cavity). The neutrophil migration was evaluated 4 h later. The

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values are means \pm SEM of six mice per group. P < 0.05, compared with carrageenan group and P < 0.05 compared with saline group (analysis of variance followed by Bonferroni's t-test).

The role played by phytoconstituents on meristematic cell is similar to that played on human cells [47]. Secondary metabolites are generated as a result of homeostasis in plants or they could the byproducts of various metabolic/ biochemical processes within the cell including primary metabolites [48]. Oleandrin, odoroside, indirubin, wrightial, carrindone, carrisol etc are very important leads from this family [49-56].

CONCLUSION

The plant N.indicum is a rich source of phytoconstituents like; cardiac glycoside, alkaloids, tannins, terpenoids, irridoids, secoirridoids and flavonoids. From the current study it is observed and hence conluded that Nerium indicum belonging to Apocynaceae family possessed anti-inflammatory effects in carrageenan-induced rat paw edema, Cotton pellet Granuloma and carrageenan induced peritonitis models, which are comparable to Diclofenac. The anti-inflammatory mechanism of the extracts may be due to the reduction of inflammatory mediators or inhibition of leukocyte rolling and neutrophil adhesion. Our Results provide new perspectives on the therapeutic use N.indicum extract and its isolated compounds in the management of inflammatory process in the body. Acknowledgement

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CONFLICT OF INTEREST- none

REFERENCES

- White, M. Mediators of inflammation and inflammatory process. Journal of Allergy and Clinical Immunology; 1999, 103: 378–381.
- Mohammed MS, Osman WJA, Garelnabi EAE, Zuheir Osman, Bashier Osman, Hassan S. Khalid, Magdi A. Mohamed. Secondary metabolites as antiinflammatory agents. The Journal of Phytopharmacology 2014; 3(4): 275-285.

- B. Nadkarni, A. K. and Nadkarni, K. M. Indian Materia Medica, Vol. I, Popular Prakashan, Bombay 1976.
- 4. Pandey Gyanendra. Materia Medica-Vegtable Drugs. In: Gyanendra P, editor. Dravyaguna Vijnana. first. Varanasi: Krishnadas Academy; 2001. p. 141–7, 123–7, 370–80.
- Prajapati ND, Purohit SS, Sharma AK, Kumar T. A Handbook of Medicinal Plants: a complete source book. 2003.Agrobios(India), Jodhpur, India. P 115carissa, nerium-361, wrightia-548.
- Warrier P K, Nambiar V P K, Ramankutty C. India Medicinal Plants: a compendium of 500 species. In: vaidya ratnam's PS V, editor. Arya Vaidya sala. first, Vol. Kottakkal: universities press; 2010. p. 386, 126, 417
- Kirtikar KR BB. Apocynaceae. In: Kirtikar KR BB and AI, editor. Indian Medicinal Plants. Second Vol II. Allahabad: Lalit Mohan Basu, Allahabad; 1935. p. 1546–9, 1581–4, 1584–6.
- 8. Abdou RH, Basha WA, Khalil WF. Subacute toxicity of Nerium oleander ethanolic extract in mice. Toxicological Research. 2019;35(3):233–9.
- Ayaz M, Yanardag S, Akkoca A, Çiçek F. Restorative Effect of Distillated Nerium Oleander Extract on Diabetic Neuropathy: Animal Model Study. Journal of Advanced Neuroscience Research. 2015; 2(2):16–21.
- Benson KF, Newman RA, Jensen GS. Antioxidant, anti-inflammatory, anti-apoptotic, and skin regenerative properties of an Aloe vera-based extract of Nerium oleander leaves (NAE-8*). Clinical, Cosmetic and Investigational Dermatology. 2015; 8:239–48.
- Chauhan S, Singh M, Thakur A, Dogra MS. Antibacterial activity of Nerium Indicum against some Gram-Positive Bacterial Species. International Journal of Drug Research and Technology. 2013;3(1):8–11.
- Dey P, Chaudhuri TK. Immunomodulatory activity of Nerium indicum through inhibition of nitric oxide and cyclooxygenase activity and modulation of TH1/TH2 cytokine balance in murine splenic lymphocytes. Cytotechnology. 2016; 68(4):749–61.
- Dey P, Chaudhuri TK. Pharmacological aspects of Nerium indicum Mill: A comprehensive review. Pharmacogn Rev. 2014; 8(16):156-162. doi:10.4103/0973-7847.134250
- 14. Dey P, Saha MR, Roy Choudhuri S, Sarkar I, Halder B, Poddar-Sarkar M, et al. Oleander Stem and Root Standardized Extracts Mitigate Acute Hyperglycaemia by Limiting Systemic Oxidative Stress Response in Diabetic Mice. Advances in Pharmacological and Pharmaceutical Sciences. vol. 2019, Article ID 7865359, 12 pages; https://doi.org/10.1155/2019/78653592019.
- 15. Fu L, Zhang S, Li N, Wang J, Zhao M, Sakai J, et al. Three new triterpenes from Nerium oleander and biological activity of the isolated compounds. Journal of Natural Products. 2005;68(2):198–206.
- Gayathri V., Ananthi S., Chandronitha C., Ramakrishnan G. S, Oleander RL& VHR. Cardioprotective effect of Nerium In, flower against



- isoproterenol-induced myocardial oxidative stress in experimental rats. J Cardiovasc Pharmacol Ther. 2011;16(1):96–104.
- 17. Ghate NB, Chaudhuri D, Panja S, Mandal N. Nerium indicum leaf alleviates iron-induced oxidative stress and hepatic injury in mice. Pharmaceutical Biology. 2015;53(7):1066–74.
- K.P. Sreena, A. Poongothai , K. Sreejith , M. Uthiralingam and S. Annapoorani. In Vitro Radical Scavenging Efficacy of different organic extracts Of Nerium indicum leaves. Pharmacologyonline. 2011; 1:155-162.
- Ke Hu, Qin Liu, Shunchun Wang KD. New oligosaccharides prepared by acid hydrolysis of the polysaccharides from Nerium indicum Mill and their anti-angiogenesis activities. Carbohydrate Research. 2009;344(2):198–203.
- Kiran C, Prasad DN. A review on: Nerium oleander Linn. (Kaner). International Journal of Pharmacognosy and Phytochemical Research. 2014;6(3):593–7.
- 21. Kumar Gaurav Singhal, Ghanshyam Das Gupta. Hepatoprotective and antioxidant activity of methanolic extract of flowers of Nerium oleander against CCl4-induced liver injury in rats. Asian Pacific Journal of Tropical Medicine (2012);677-685.
- Lingpeng Dai, Wanxian Wang, Xinjiao Dong, Renyong Hu XN. Molluscicidal activity of cardiac glycosides from Nerium indicum against Pomacea canaliculata and its implications for the mechanisms of toxicity. Environmental Toxicology and Pharmacology. 2011;32(2):226–32.
- Luay J. Rashana, Katrin Franke, Myint Myint Khinea, Gerhard Kelter, Heinz H. Fiebig, Joachim Neumann LAW. Characterization of the anticancer properties of monoglycosidic cardenolides isolated from Nerium oleander and Streptocaulon tomentosum. Journal of Ethnopharmacology. 2011; 134:781–8.
- Mouhcine M, Amin L, Saaid A, Khalil H, Laila B, Mohammed EM. Cytotoxic, antioxidant and antimicrobial activities of Nerium oleander collected in Morocco. Asian Pacific Journal of Tropical Medicine. 2019;12(1):32–7.
- Ni D., Madden T.L., Johansen M., Felix E. HDH& NRA. Murine pharmacokinetics and metabolism of oleandrin, a cytotoxic component of Nerium oleander. J Exp Ther Oncol. 2002;2(5):278–85.
- Pathak, S; Multani, AS; Narayan, S; Kumar, V; Newman R. AnvirzelTM, an extract of Nerium oleander, induces cell death in human but not murine cancer cells. Anticancer Drugs. 2000;11(6):455–63.
- Ray SD, Ray S, Zia-Ul-Haq M, De Feo V, Dewanjee S. Pharmacological basis of the use of the root bark of Zizyphus nummularia Aubrev. (Rhamnaceae) as antiinflammatory agent. BMC Complement Altern Med. 2015 Nov 23;15:416. doi: 10.1186/s12906-015-0942-7. PMID: 26597878; PMCID: PMC4657250.
- Ismail TS, Gopalakrishnan S, Begum VH, Elango V. Anti-inflammatory activity of Salacia oblonga Wall.

- and Azima tetracantha Lam. J Ethnopharmacol 1997; 56:145–52.
- M.R. Sulaiman, E.K. Perimal, M.N. Akhtar , A.S. Mohamad, M.H. Khalid, N.A. Tasrip a,F. Mokhtar, Z.A. Zakaria, N.H. Lajis, D.A. Israf. Anti-inflammatory effect of zerumbone on acute and chronic inflammation models in mice. Fitoterapia 81 (2010) 855–858.
- Raveen R, Pandeeswari M, Ahmed F, Tennyson S, Arivoli S. Bioefficacy of Nerium oleander Linnaeus (Apocynaceae) floral extracts on the larva of three vector mosquitoes of medical importance. International Journal of Mosquito Research. 2017;4(6):65–77.
- 31. Sabira Begum, Razia Sultana, Bina S. Siddiqui. Triterpenoids from the leaves of Nerium oleander.Phytochemistry, Volume 44, Issue 2, January 1997, Pages 329-332.
- 32. Sharma pallavi AC. Chemical Constituents of Plants from the Genus Nerium. Chemistry and Biodiversity. 2010;1198–207.
- 33. Sharma, P., Gupta, Y.K., Sharma, M.C., Dhobal, M.P., "Two new compounds from the stem of Nerium oleander" Indian Journal of Chemistry 2012, 49B, 374-378.
- 34. Siddiqui B.S., Sultana R., Begum S. ZA& SA. Cardenolides from the methanolic extract of Nerium oleander leaves possessing, Central nervous system depressant activity in mice. J Nat Prod. 1997;60(6):540–4.
- 35. Siddiqui S, Hafeez F, Begum S, Siddiqui BS. Oleanderol, a new pentacyclic triterpene from the leaves of nerium oleander. Journal of Natural Products. 1988;51(2):229–33.
- Siddiqui, B.S., Khatoon, N., Begum, S., Farooq, A. D., Qamar, K., Bhatti, H.A., Ali, S.K., "Flavonoid and cardenolide glycosides and a pentacyclic triterpene from the leaves of Nerium oleander and evaluation of cytotoxicity." Phytochemistry 2012, 77, 238-244.
- 37. Siddiqui, S., Hafeez, F., Begum, S., Siddiqui, B.S., "Isolation and structure of two cardiac glycosides from the leaves of Nerium oleander" Phytochemistry 1987, 26(1), 237-241
- 38. Siddiqui, S., Hafeez, F., Begum, S., Siddiqui, B.S., "Two triterpenes from the leaves of Nerium oleander" Phytochemistry1989, 28(4), 1187-1191.
- 39. Singhal KG, Gupta G Das. Hepatoprotective and antioxidant activity of methanolic extract of flowers of Nerium oleander against CCI 4-induced liver injury in rats. Asian Pacific Journal of Tropical Medicine. 2012;5(9):677–85.
- 40. Smith JA, Madden T, Vijjeswarapu M, Newman RA. Inhibition of export of fibroblast growth factor-2 (FGF-2) from the prostate cancer cell lines PC3 and DU145 by Anvirzel and its cardiac glycoside component, oleandrin. Biochem Pharmacol 2001; 62(4): 469-472
- 41. Tirumalasetti J, Patel M, Shaikh U, Harini K, Shankar J. Evaluation of skeletal muscle relaxant activity of aqueous extract of Nerium oleander flowers in



- Albino rats. Indian Journal of Pharmacology. 2015;47(4):409–13.
- 42. Turan N, Akgün-Dar K, Kuruca SE, Kiliçaslan-Ayna T, Seyhan VG, Atasever B, et al. Cytotoxic effects of leaf, stem and root extracts of Nerium oleander on leukemia cell lines and role of the p-glycoprotein in this effect. J Exp Ther Oncol 2006; 6(1): 31-38.
- 43. Vijayvergia R, Kumar J. Quantification of Primary Metabolites of Nerium indicum Mill Introduction: Materials and Method: Estimation of carbohydrates: 2007;21(1):123–8.
- 44. Yong P, Patrea R, Lin T, Carrie C, Ho-Jeong L, Murali KR, et al. PBI-05204, a supercritical CO2 extract of Nerium oleander, inhibits growth of human pancreatic cancer via targeting the PI3K/mTOR pathway. Invest New Drugs 2015; 33(2): 271-279
- 45. Zhao M, Bai L, Wang L, Toki A, Hasegawa T, Kikuchi M, et al. Bioactive cardenolides from the stems and twigs of Nerium oleander. Journal of Natural Products. 2007;70(7):1098–103.
- 46. Sautebin L, Ialenti A, Ianaro A, Di Rosa M. Endogenous nitric oxide increases prostaglandin biosynthesis in carrageenin rat paw oedema. Eur J Pharmacol 1995; 286: 219-222.
- Salvemini D, Wang ZQ, Wyatt PS, Bourdon DM, Marino MH, Manning PT, et al. Nitric oxide: a key mediator in the early and late phase of carrageenaninduced rat paw inflammation. Br J Pharmacol 1996; 118: 829-838.
- Vogel H.G., In Drug Discovery and Evaluation, Pharmacological Assay, 2nd Edn.2002; Vogel HG, Vogel WH (Eds). Springer Verlag: Berlin, Heidelberg Germany.
- 49. Winter, C.A., Porter, C.A., 1957. Effect of alteration inside chain upon antiinflammatory and liver

- glycogen activities in hydrocortisone esters. Journal of American Pharmaceutical Association 46, 515–519
- Winter, C.A., Risely, E.A., Nuss, G.W., 1962. Carrageenen-induced edema in the hind paw of rat as an assay for antiinflammatory drugs. Proceedings of Society for Experimental Biology and Medicine 111, 544–547.
- 51. Robert Fürst, Ilse Zündorf, "Plant-Derived Anti-Inflammatory Compounds: Hopes and Disappointments regarding the Translation of Preclinical Knowledge into Clinical Progress", Mediators of Inflammation, vol. 2014, Article ID 146832, 9 pages, 2014. https://doi.org/10.1155/2014/146832
- 52. Akkol EK (2012) New Strategies for Anti-Inflammatory Drug Development. J Pharmacogenom Pharmacoproteomics 3: e118. doi:10.4172/2153-0645.1000e118
- 53. Patil KR, Mahajan UB, Unger BS, Goyal SN, Sateesh Belemkar S, Surana SJ, Ojha S, Patil CR. Animal Models of Inflammation for Screening of Antiinflammatory Drugs: Implications for the Discovery and Development of Phytopharmaceuticals Int. J. Mol. Sci. 2019, 20, 4367; doi:10.3390/ijms20184367.
- 54. Liu, X., Zheng, J. & Zhou H. TLRs as pharmacological targets for plant-derived compounds in infectious and inflammatory diseases. International Immunopharmacology. 2011; 11:1451–6.
- Khosrow Kashfi. Anti-Inflammatory Agents as Cancer Therapeutics. Advances in Pharmacology; 2009, 57: 31-89.
- 56. Corlett, R.T. Plant diversity in a changing world: Status, trends, and conservation needs. Plant. Diverse. 2016, 38, 10–16. OVEREXPLOITATION