

CADMIUM TOXICITY- A HEALTH HAZARD AND A SERIOUS ENVIRONMENTAL PROBLEM -AN OVERVIEW

V.PRAVEEN CHAKRAVARTHI, J.A. PRADEEP KIRAN AND M.BHASKAR*

*Division of Animal Biotechnology, Department of Zoology,
Sri Venkateswara University, Tirupati - 517502, AP, India.*

*Corresponding Author Email: matchabhaskar2010@gmail.com

ABSTRACT

Cd^{2+} a heavy metal accumulates in the body and has a very long biological half-life (10–30 years) in humans. Environmental cadmium contamination occurs through industries related to batteries, coatings, electroplating, alloys, plastics, glasses, ceramics, enamels and artists' colors, zinc, copper, lead, and iron ores treatment, coal and oil burning power plants, weathering of parent rocks etc. This review is mainly focused on general characteristics of cadmium, effect on human health in relation to different organs such as kidneys, liver, lungs, reproductive system etc. and mechanism of toxicity. Renal toxicity, hepatotoxicity and carcinogenicity are the major effects of cadmium induced toxicity. Production of reactive oxygen species, free radical induced cell damage, interference with calcium and vitamin-D metabolism (major mechanism involved in cadmium induced bone damage), oxidative DNA damage, glutathione depletion, accumulation as metallothionein complex (Renal toxicity), CFTR (Cystic fibrosis transmembrane conductance regulator) dysfunction (lung toxicity), chromosomal aberrations, modification of transcription factors, altering the antioxidant defense system are the observed mechanisms involved in cadmium induced cytotoxicity. Exogenous supplementation of glutathione (GSH) and metallothionein (MT) will play protective roles against cadmium toxicity. Reducing the use of cadmium in industries or avoiding of Cadmium exposure and recycling the cadmium products prevent the cadmium induced cytotoxicity to some extent.

KEYWORDS

Cadmium, Cytotoxicity, Genotoxicity, Environmental pollution, Kidney, Liver.

INTRODUCTION

Cadmium (Cd^{2+}) is one of the most common nonessential elements, relatively accessible heavy metal in our environment causing wide range of toxic effects. Cd^{2+} accumulates in the body and has a very long biological half-life (10–30 years) in humans¹⁻³. The most common forms of cadmium found in the environment exist in combinations with other elements such as cadmium oxide, cadmium chloride and cadmium sulfide etc. Inhalation and accumulation of cadmium disturbs various metabolic functions and there by affects the human health. Effect of cadmium on various organs of the human body

has been studied earlier and several mechanisms have been proposed. In this review all those mechanisms have been scrutinized to some extent and propose some pathways for cadmium induced toxicity on various organs of the human body.

Toxic effects of Cadmium:

Impact of Cadmium on Human health

Exposure to cadmium through food is typical for most people but is not a major health concern. This is because the cadmium present in the body from our diet is about 0.0004mg/kg/day. This figure is about ten times lower than the level of

cadmium that causes kidney damage from eating contaminated food (Table.1).

Table.1 Cadmium levels in Environment

| Natural cadmium levels in the environment | |
|---|------------------------------|
| Atmosphere | 0.1 to 5 ng/m ³ * |
| Earth's crust | 0.1 to 0.5 µg/g** |
| Marine sediment | ~1 µg/g |
| Sea-water | ~0.1 µg/l*** |

*ng/m³ = nanograms (10E-9 g) per cubic meter.

** µg/g = micrograms (10E-6 g) per gram.

*** µg/l = micrograms (10E-6 g) per liter.

Cadmium exposure may be implicated in some humans disorders related to hyper activity and increased aggressiveness⁴. Cadmium can enter to body by inhalation and other routes and accumulates mainly in the kidneys. At high levels, it can reach a critical threshold and can lead to serious kidney failure. Cadmium can enter through ingestion, intraperitoneal, subcutaneous, intramuscular and intravenous routes Vahter et al⁵ that Cd²⁺ retention is generally higher in women than in men. "Ouch-ouch" or *itai-itai* is mainly a women disease. This disease is caused by long-term exposure of the inhabitants of Tayoma in Japan to Cd²⁺ intoxication. Clinical features of this disease include renal tubular dysfunction, osteomalacia, amino-aciduria, glycosuria and anemia which include decreased Hb, iron deficiency and low serum erythropoietin levels⁶ in the human Hotz et al⁷ shown that kidney effects may be reversible at low exposures once cadmium exposure is reduced or removed.

Cadmium – Carcinogenesis:

Cadmium affects cell proliferation, differentiation, apoptosis and other cellular activities and can cause numerous molecular lesions' that would be relevant to carcinogenesis. For a long time cadmium has been considered as a non-genotoxic carcinogen, as it is only weakly mutagenic in bacterial and mammalian cell test systems. Recently, it was evidenced that when assayed in a

test system, in which both intragenic and multilocus mutations can be detected, cadmium acts as a strong mutagen which induces predominantly multilocus deletions. It was proposed that two mechanisms might play an important role in cadmium induced mutagenicity. They were 1.Induction of reactive oxygen species (ROS); and 2.Inhibition of DNA repair (**Figure.1**). Experimental evidence suggests that cadmium at relevant concentrations induces mutations by oxidative DNA damage and that it decreases genetic stability by inhibiting the repair of endogenous and exogenous DNA lesions, which in turn increase the probability of mutations and consequently cancer initiation by this metal. However, cadmium at nontoxic doses interferes with DNA repair processes and enhances the genotoxicity of directly acting mutagens.

Carcinogenic effect of cadmium in humans and in experiments with rats was discussed and concluded that, in rats induction of cancer in different organs by cadmium exposure was conformed but whereas in humans yet further studies are needed to conform the carcinogenic effect⁸. The rat testis may also develop tumors if cadmium is given peritoneal at high doses. Subsequent to testicular hemorrhagic necrosis, there will be loss of testosterone production and hyperplasia and neoplasia of testicular interstitial

cells, thought to be a response to trophic hormone release from the pituitary.

Regarding the apparent discrepancy between the results of human studies and those of laboratory animal studies, it should be clarified whether common mechanisms for the occurrence of

carcinogenicity exist, and the apparent discrepancy should be explained at the molecular and cellular levels. Which means "The agent is probably carcinogenic to humans" based on the existing evidence proving it as causative for carcinogenesis.

Fig. 1: Showing Mechanisms involved in cadmium induced mutagenicity.

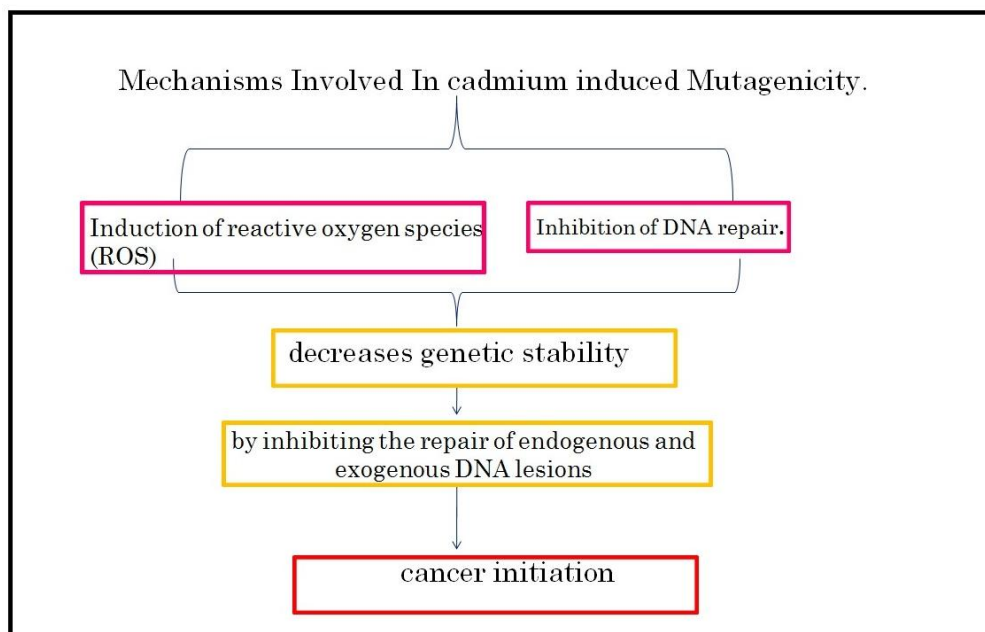
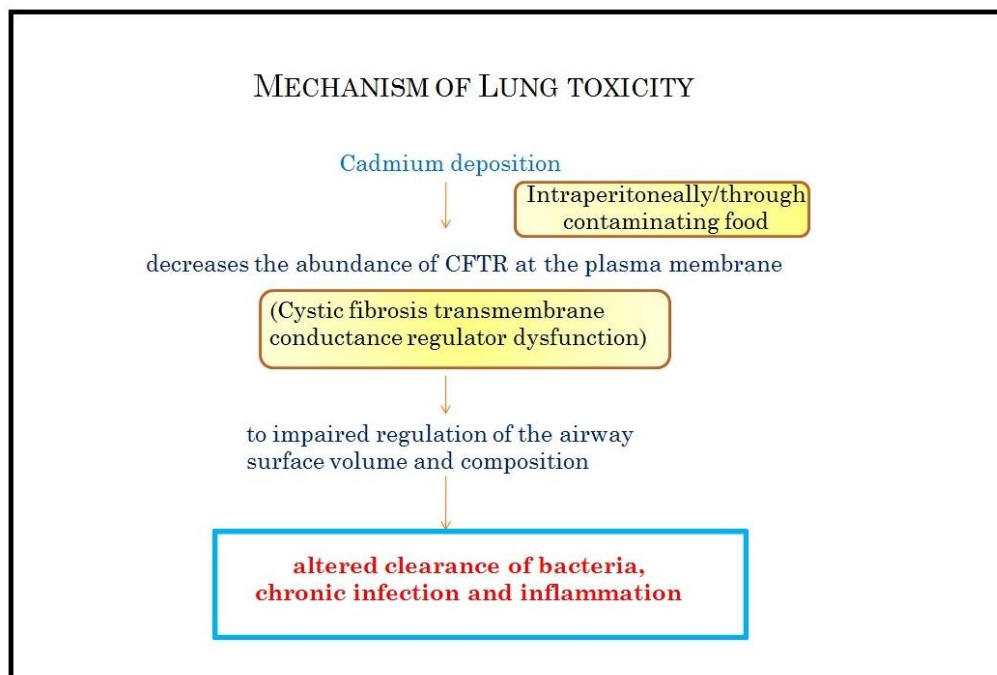


Fig. 2: Showing mechanism involved in cadmium induced Lung toxicity.



Genotoxic effect of Cadmium

Cadmium exposure has been established to induce cancer in various tissues of laboratory animals. Contrary to early findings of the lack of genotoxicity by cadmium, recent findings of mammalian cell culture studies have revealed genotoxic effects. Furthermore, cadmium exposure at relatively low doses induces circulatory diseases in laboratory animals. Despite such results of various cadmium toxicities in animal studies, data from human studies are lacking and insufficient to support the cause-effect relationship.

Non-genotoxic mechanisms upregulating intracellular signaling pathways leading to increased mitogenesis are discussed as major mechanisms for the interpretation of the carcinogenic activity by chronic cadmium exposure. About 1 μ M cadmium stimulates DNA synthesis and cell proliferation in various cell lines, whereas more elevated concentrations are inhibitory. It stimulates the expression of immediate early genes (c-fos, c-jun, and c-myc), of the tumor suppressor gene p53, and of genes coding for the syntheses of protective molecules, including metallothioneins, glutathione, and stress (heat shock) proteins. The mechanisms underlying the modulation of gene activity by cadmium are discussed in terms of interference with cellular signaling at the levels of cell surface receptors, cellular calcium and zinc homeostasis, protein phosphorylation, and modification of transcription factors. In considering the available evidence, the carcinogenic properties of cadmium were interpreted using a multifactorial approach involving indirect genotoxicity (interference with DNA repair) and the up regulation of mitogenic signaling pathways.

Impact of cadmium on cell signaling pathways and in the induction of apoptosis:

The effect of Cd in altering the different signaling pathways and in induction of apoptosis^{9, 10},

Showed Cd induced apoptosis in different cell lines of different organisms including humans and reported that unlike Fenton-type metals Cd cannot produce reactive oxygen species directly; the apoptotic effects of cadmium at least in part are mediated via induction of oxidative stress. In C6 rat glioma cells, cadmium caused externalization of phosphatidylserine, break down of the mitochondrial membrane potential, and activation of caspase-9; inter nucleosomal DNA fragmentation, chromatin condensation and nuclear fragmentation. Cadmium as similar to H₂O₂ is a potent inducer of apoptosis in C6 cells¹¹. Cd-induced apoptosis is partly caused by caspase-9 activation triggered by Cytc¹². Cadmium caused the PTP opening possibly through its binding to thiol groups of ANT. Furthermore, the mechanism of the PTP opening induced by cadmium was probably distinct from that of the calcium-induced PTP opening¹³. The apoptotic effects of cadmium at least in part are mediated via induction of oxidative stress¹¹.

A rapid and transient ROS generation by Cd triggers apoptosis via caspase-dependent pathway and subsequent mitochondrial pathway. CdCl₄ treatment significantly increased the levels of apoptotic proteins such as caspases-3, PARP, Bax, Bid and cytochrome C and also increased the levels of inflammatory mediator's iNos and Cox-2¹⁴.

Cadmium – Cardiovascular dysfunction:

The incidence of cardiovascular disease has increased in the general population, and cardiac damage is indicated as one important cause of mortality. Recent investigation has established that free radicals may be important contributors to cardiac dysfunction and myocardial damage¹⁵. In our laboratory¹⁶ reported that the cadmium toxicity could have induced oxidative damage in both liver and kidney by enhancing peroxidation of membrane lipids due to inhibition of the antioxidant enzymes,¹⁷ has been carried some

investigations in our laboratory and reported cadmium toxicity induced alterations in heart and muscle tissues of rabbit leading to changes in blood constituents, abnormalities in heart and muscle function altering glycolysis, citric acid cycle, phosphatase metabolism, transamination reactions and induction of free radical stress. The cardiovascular effect of cadmium was discussed by ⁹ and analyzed that renal tubular dysfunction by cadmium was associated with regulation of blood pressure.

Since both cadmium exposure and the incidence of cardiac damage have increased in recent years, Cadmium exposure could initiate some series of events that occur in the heart and result in the production of free radicals. If so, free radicals might contribute to the alterations processes in heart which result in further injury. Cd induced atherosclerosis was reported in rabbit ¹⁸. According to ¹⁹ pretreatment with zinc produces tolerance to several cadmium toxic effects. They also reported that zinc-induced protection against the cytotoxicity of cadmium in stellate cells may be related to the maintenance of normal redox balance inside the cell. Thus the free radical induced damage was found to be major mechanism of cadmium induced cardiovascular diseases.

Cadmium – Lung damage:

The lungs absorb cadmium more efficiently than the gastrointestinal tract ²⁰. Takenaka et al ²¹ that inhaled ultrafine Cd²⁺ oxide particles cloud cause lung injury in rats. An association between cadmium exposure and an increased risk of pulmonary diseases such as lung cancer and chronic obstructive pulmonary disease has been reported in humans and animals ²². Cd induced CFTR (Cystic fibrosis transmembrane conductance regulator) dysfunction leads to impaired regulation of the airway surface volume and composition resulting in altered clearance of bacteria and chronic infection and

inflammation ²³. Cadmium decreases the abundance of CFTR at the plasma membrane resulting in a decrease in chloride transport in epithelial cells present in the lung ²⁴. Minute amount of cadmium deposit in lungs if administered intraperitoneal or through contaminating food, it can still induces inflammation and proliferation due to persistent presence in lung cells but these two events may occur independently ²⁵. Thus the toxic effect of cadmium on lungs was mainly due to CFTR dysfunction, which results in altered bacterial clearance, there by infection and inflammation (Figure.2).

Cadmium – Hepatotoxicity:

The liver is the primary target organ following acute systemic Cd exposure. Approximately half of Cd absorbed systemically is rapidly accumulated in the liver, which results in the reduced availability of Cd to such organs as the kidneys and testes, which are more sensitive to its toxic actions ²⁶. Productions of reactive oxygen species and oxidative tissue damage due to Cd have been associated with hepatotoxicity. It has been demonstrated that Cd produces both dose- and time-dependent increases in intracellular glutathione concentration during chronic environmental or occupational exposure at low doses. However, with high level acute Cd exposure, significant glutathione depletion occurs. Moreover, Cd is known to cause a reduction in glutathione content in isolated hepatocytes ²⁷. Liver injury due to acute Cd exposure is dominated by apoptosis and necrosis. Apoptosis seems to play a major role in eliminating damaged cells and its participation is profound in nonparenchymal liver cells where it represents the major type of cell death ²⁸.

Histological evaluation of liver injury reveals that acute toxicity comprises hepatocellular swelling, sinusoidal congestion, pyknosis, and karyorrhexis ²⁹. Cd induced hepatotoxicity has

been shown to cause early cellular changes in the rough endoplasmic reticulum and nucleus³⁰. Later alterations include mitochondrial swelling and the appearance of fibrillar material within the cytoplasm³¹.

Several reports have addressed the direct mechanism of Cd-induced hepatotoxicity at the sub-cellular and molecular level. Sub-cellular localization of Cd demonstrates that Cd is distributed to the nucleus mitochondria and endoplasmic reticulum, which localizes Cd in target organelles³². Furthermore, Cd administration may also bring about DNA damage³³. CdCl₄ significantly increased the levels of lipid peroxides, oxidized glutathione and decreased the levels of reduced glutathione, SOD (Superoxide dismutase) and CAT (catalase)¹⁴. Production of reactive oxygen species, oxidative tissue damage, apoptosis, glutathione depletion were found to be the major mechanisms attributed for cadmium induced liver injury.

Renal toxic Effects:

Cadmium can enter to body by inhalation and other routes and accumulates mainly in the kidneys. At high levels, it can reach a critical threshold and can lead to serious kidney failure. Cadmium can enter through ingestion, intraperitoneal, subcutaneous, intramuscular and intravenous routes. Expression of the ZIP8 metal ion transporter (*Slc39a8* gene) appears to be a key factor contributing to the selective toxicity of Cd in the endothelial cells of organs such as the testes and kidneys³⁴⁻³⁵. Cadmium and lead as chronic kidney disease risk factors in the general population and provide novel evidence of risk with environmental exposure to both metals. Severity of renal toxicity increases with Cd accumulation in the kidney, which depends on the level and duration of exposure. A very important finding of this study is that Cd affects the kidney, especially the main tubules,

even at relatively low accumulation levels in this organ.

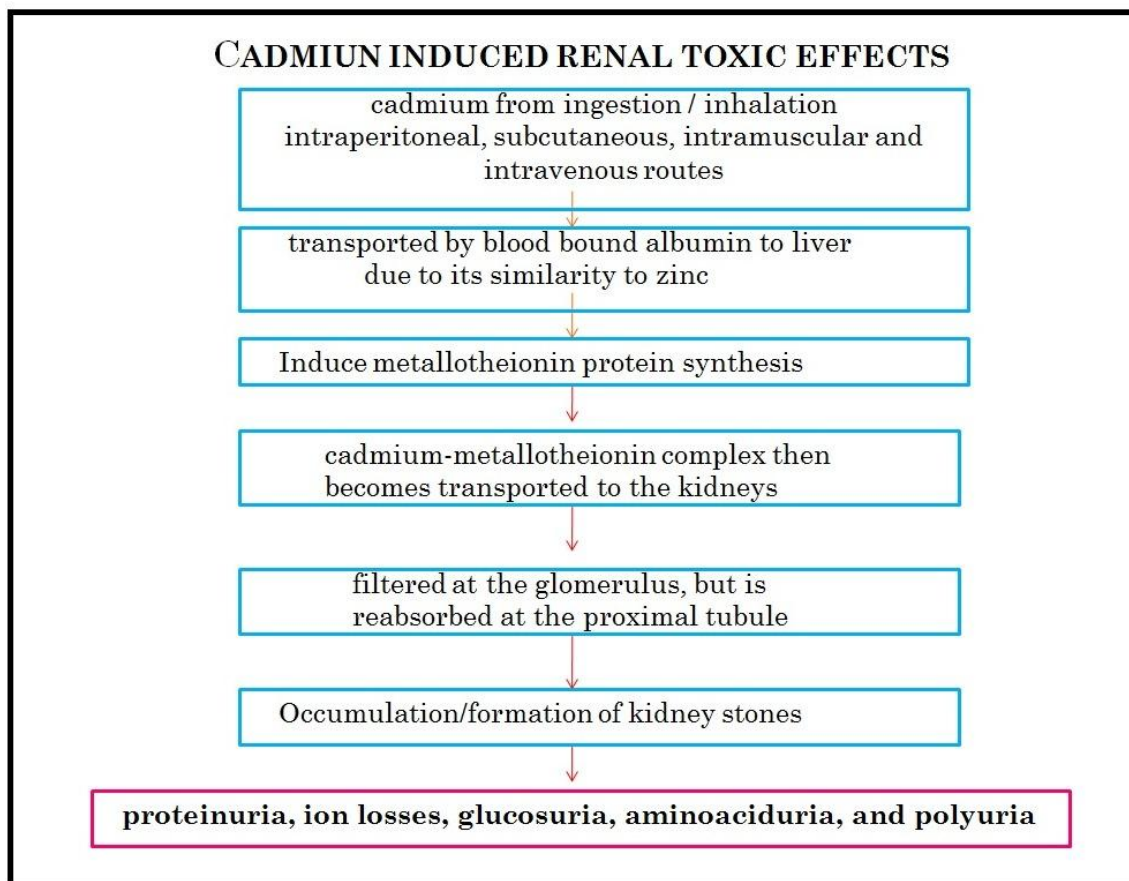
The lower and long-term exposure to cadmium through air or through diet can cause kidney damage. Although the damage is not life-threatening, it can lead to the formation of kidney stones and affect the skeleton, which can be painful and debilitating. Humans have a daily intake of cadmium from ingestion and inhalation which is around 20 to 40 µg per day, but only 5 to 10% of this is absorbed. After absorption, cadmium is transported into the blood bound albumin. It is taken up by the liver and due to its similarity to zinc, causes this organ to induce the synthesis of the protein metallothionein (MT) to which it binds. The cadmium-metallothionein complex then becomes transported to the kidneys, and it is filtered at the glomerulus, but is reabsorbed at the proximal tubule where it remains stored therefore; the kidney is one of the main target organs for cadmium induced toxicity [36]. Within the renal tubular cells, the cadmium-MT complex becomes degraded by digestive enzymes, which releases the cadmium. Renal tubular cells deal with the release of this toxic substance by synthesizing MT to neutralize it, but eventually the kidneys lose their synthetic capacity for MT. At this point, the cadmium has accumulated to a high level in the renal tubular cells, and irreversible cell damage occurs (Figure.3)³⁷.

The renal toxicity of cadmium in humans and in various experimental animal studies from the previous studies and concluded that induction of renal dysfunction may be partly dependent upon the biosynthesized amounts of metallothioneins in the kidney⁹. Several mechanisms have been proposed to explain the toxic effect of Cd²⁺ on renal cells. Cd may cause nephrotoxicity by generating free radicals³⁸⁻³⁹. Interestingly, a protective effect of zinc (Zn²⁺) has been reported in vitro against the cellular toxicity due to Cd²⁺.

Zn²⁺ protection is probably due to an action on oxidative stress and apoptosis⁴⁰ while Cd²⁺ induces cytochrome C release from

mitochondria, leading to apoptosis via the activation of the caspase 3 and 9 cascade (Figure.3)⁴¹.

Fig. 3: Showing cadmium induced renal toxicity



Cadmium – bone metabolism:

Bone density and biomarkers such as parathyroid hormone, calcitonin and bone specific alkaline phosphatase activity levels were studied on exposure to Cd in women population⁴².

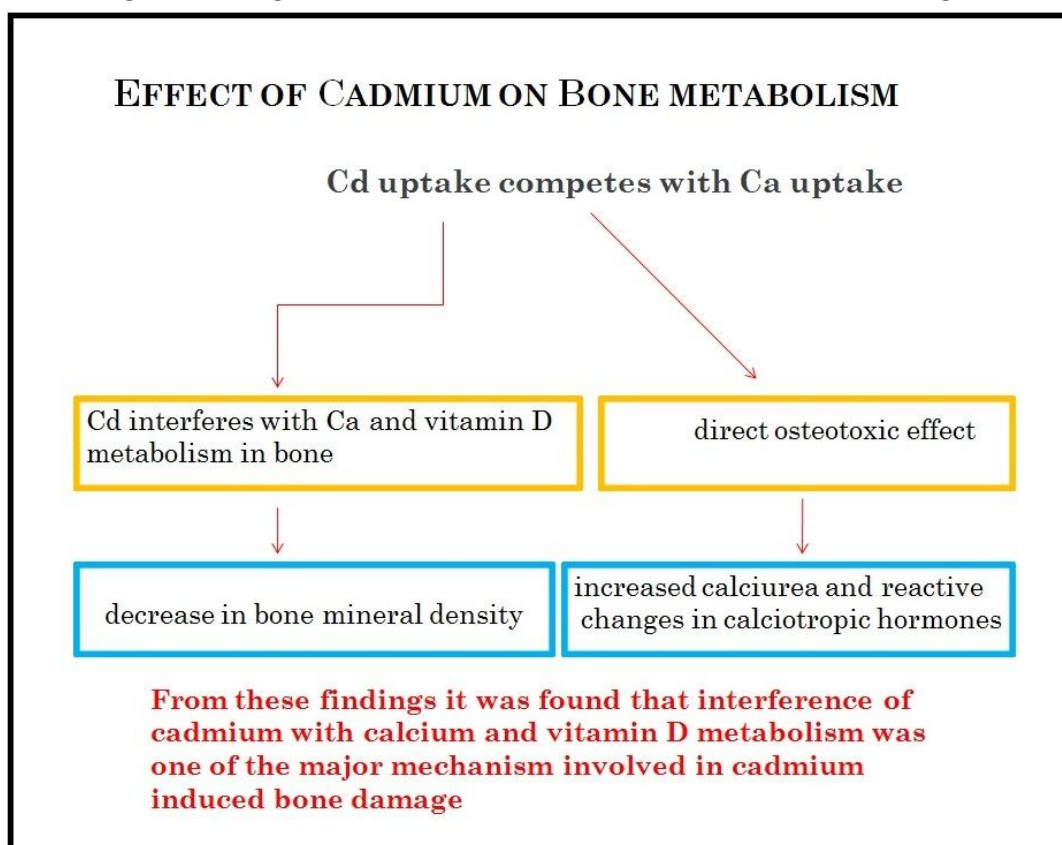
Renal dysfunction and bone metabolism disorder are known to be the representative adverse health effects of chronic Cd toxicity in humans and animals. Other effects of Cd exposure on humans and animals are also recognized as follows: cases of acute and chronic toxicity due to occupational and environmental exposure, such as anemia, respiratory disorder,

hypertensive and cardiovascular effects, nervous system symptoms, cancer of the lung and prostate, toxic effects on the placenta and teratogenicity^{43, 44}. Particularly, Cd interferes with Ca and vitamin-D metabolism in bone, kidney and intestine. The interaction between Cd and Ca in bone, intestine, and kidney may result in the disorder of bone metabolism. On the other hand, pregnancy and lactation are also important physiological factors affecting bone metabolism in the mother. Ca absorption is decreased by competition with Cd in the intestine, and more Ca is released from maternal bone and transferred to neonate by lactation. In

the intestine, Cd uptake competes with Ca uptake. Cd causes a marked decrease in bone density compared to the normal decrease in bone mineral density during lactation. Lactation is an important factor contributing to the decrease in bone mineral density and Cd has an additive effect of decreasing bone metabolism of mother animal, although the Cd intake level is relatively low (approximately 3-14µg Cd/kg/day). Environmental exposure to cadmium increases bone resorption in women, suggesting a direct osteotoxic effect with increased calciurea and reactive changes in calciotropic hormones⁴². Long-term dietary exposures in rats, at levels corresponding to environmental exposures in

humans, result in increased skeletal fragility and decreased mineral density. Cadmium-induced demineralization begins soon after exposure, within 24 hours of an oral dose to mice. In bone culture systems, cadmium at low concentrations acts directly on bone cells to cause both decreases in bone formation and increases in bone resorption, independent of its effects on kidney, intestine, or circulating hormone concentrations⁴⁵. Cd administration affects bone marrow, spleen and thymus (Figure.4)⁴⁶. From these findings it was found that interference of cadmium with calcium and vitamin-D metabolism was one of the major mechanisms involved in cadmium induced bone damage.

Fig. 4: Showing mechanism involved in cadmium induced bone damage



Effect on reproduction:

Cd²⁺ has been shown to exert significant effects on ovarian and reproductive tract morphology,

with extremely low dosages reported to simultaneous ovarian luteal progesterone biosynthesis and high dosages inhibiting it⁴⁷.

Survival of adult insect *Oncopeltus fasciatus* females was decreased at concentrations higher than 10 mg Cd/l, while males were only affected at 30 mg Cd/l or higher doses. Reproduction was the most affected parameter. Oviposition rate, fecundity and fertility of females exposed to 10 mg Cd/l were significantly lower than controls (73%, 58% and 55% relative to controls, respectively). A reduction in adult survival and no reproduction at the highest concentrations of Cd and Zn was observed in *Proisotomaminuta* Tullberg (Collembola)⁴⁸.

Cadmium is directly toxic to primary Leydig cells, and that the decreased percentage of normal cells and the increased level of DNA damage in cadmium-exposed Leydig cells may be responsible for decreased testosterone secretion in rats⁴⁹.

The combined exposure of lead and Cd²⁺ causes decreased glutathione status and SOD activities in rat ovarian granulosa cells.⁵⁰ They reported that these toxic metals disturb membrane integrity of cells via reactive oxygen species (ROS) and thereby classifying mechanism for altered receptor binding, steroidogenesis, and hormone production⁵¹, observed increased Cd²⁺ concentrations and shortened prothrombin time leads to a stage of hyper coagulation which in turn leads to a risk of thrombosis in rats. Significant positive association between the percentage of immotile sperms and seminal plasma levels of lead and cadmium was observed⁵². No association is indicated for blood cadmium from women and oocyte fertilization, adjusted for urine cadmium, however, an inverse adjusted association between blood cadmium from men and oocyte fertilization (relative risk=0.66, P=0.143) was observed⁵³.

In SPEED 98 (Strategic Programs on Environmental Endocrine Disruptors '98) declared that cadmium is one of the 70 susceptible elements found to have adverse

effects on endocrine system. Production of Reactive oxygen species, necrosis, and inhibition of progesterone biosynthesis, decreased glutathione status and SOD activities in were the so far observed mechanisms of cadmium induced reproductive toxicity.

CONCLUSION

Finally it is concluded that, based on existing literature a significant association was observed between risk of pathogenicity and environmental exposure to cadmium. An attempt has been made first time to project such an association to our knowledge has been reported in relation to cadmium toxicity. These findings of information helps to understand the cadmium induced serious health hazard leading to necessitating the importance of preventive measure formulation and practice.

REFERENCE

1. Friberg L, Piscator M, Nordberg GF and Kjellstrom T. *Cadmium in the environment*. 2nd edition. Cleveland, CRC Press ;(1974).
2. Goyer RA. *Nutrition and Metal Toxicity*. Am J Clin Nutr, 61: 646S-650S, (1995).
3. Jarup L, Berglund M, Elinder CG, Nordberg G, Vahter M. *Health effects of cadmium exposure – a review of the literature and a risk estimate*. Scand J work Environ Health, 24: 1-51, (1998).
4. Lopez E, Arce C, Oset-Gasque MJ, Canadas S, Gonzalez MP. *Cadmium induces reactive oxygen species generation and lipid peroxidation in cortical neurons in culture*. Free Radic Biol Med; 40:940–51, (2006).
5. Vahter M, Berglund M, Akesson A, Liden C. *Metals and women's health*. Environ Res, 88:145-55, (2002).
6. Tsukahara T, Ezaki T, Moriguchi J, Furuki K, Fukui Y, Ukai H, Okamoto S, Sakurai H. *No significant effect of iron deficiency on cadmium body burden or kidney dysfunction among women in the general population in Japan*. Int Arch Occup Environ Health, 76: 275-81, (2003)
7. Hotz P, Buchet JP, Bernard A, Lison D, Lauwerys R. *Renal effects of low-level environmental cadmium exposure: 5-year follow-up of a subcohort from the Cadmibel study*. Lancet, 354 (9189):1508-1513, (1999).

8. Waalkes MP, Rehm S, Riggs CW, Bare RM, Devor DE, Poirier LA, Wenk ML, Henneman JR, Balaschak MS. *Cadmium carcinogenesis in male Wistar [Cr:(WI)BR] rats: dose-response analysis of tumor induction in the prostate and testes and at the injection site.* Cancer Res; 48:4656-63, (1988).
9. Satoh M, Koyama H, Toshiyuki K, Hideaki K, Chiharu T. *Perspectives on Cadmium Toxicity Research.* J Exp Med; 196:23-32, (2002).
10. Watjen W, Monika C, Marta B, Detmar B. *Cadmium-induced apoptosis in C6 glioma cells: Mediation by caspase 9-activation.* BioMetals; 15: 15-25, (2002)
11. Watjen W and Beyersmann D. *Cadmium-induced apoptosis in C6 glioma cells: Influence of oxidative stress.* BioMetals; 17: 65-78, (2004).
12. Kondoh M, Araragi S, Sato K, Higashimoto M, Takiguchi M, Sato M. *Cadmium induces apoptosis partly via caspase-9 activation in HL-60 cells.* Toxicology; 170(1-2):111-7, (2002).
13. Min L, Tian X, Chun-Sun J, Lin-Jiang L, Juan-Ling F, Zong-Can Z. *Cadmium directly induced the opening of membrane permeability pore of mitochondria which possibly involved in cadmium-triggered apoptosis.* Toxicology; 194(1-2):19-33, (2003).
14. Srilaxmi P, Gangadhara Reddy S, Bilhan Kavikishore P, Hussainaiah Setty O, Prakash Babu P. *Protective efficacy of natansnin, a dibenzoyl glycoside from Salvinianatans against CCl4 induced oxidative stress and cellular degeneration in rat liver.* BMC Pharmacology; 10:13, (2010).
15. Pichardo J, Palace V, Farahmand F and Singal PK. *Myocardial oxidative stress changes during compensated right heart failure in rats.* Mol Cell Biochem; 196:51-57, (1999).
16. MohanaRadhika M. *Toxicological impact of cadmium on haematological and biochemical modulations in female Rabbits.* Ph.D thesis, S.V. University, Tirupati. (2007).
17. Subbarao K. *Effect of cadmium on metabolic modulations in heart and muscle of female Rabbits.* Ph.D thesis, S.V. University, Tirupati. (2008).
18. Subramanyam G, Bhaskar M and Govindappa S. *The role of Cadmium induction of Atherosclerosis in rabbits.* Ind Heart J; 44 (3): 177-180, (1992).
19. Souza V, Del M, Escobar C, Bucio L, Hernandez E, Gutierrez MC. *Zinc pretreatment prevents hepatic stellate cells from cadmium-produced oxidative damage.* Cell Biology and Toxicology; 20(4): 241-51, (2004).
20. Jin T, Nordberg M, Frech W, Dumont X, Bernard A, Ye TT, Kong Q, Wang Z, Li P, Lundström NG, Li Y, Nordberg GF. *Cadmium biomonitoring and renal dysfunction among a population environmentally exposed to cadmium from smelting in China (ChinaCad).* Biometals; 15, 397-410, (2002)
21. Takenaka S, Karg E, Kreyling WG, Lentner B, Schulz H, Ziesenis A, Schramel P, Heyder J. *Fate and Toxic Effects of Inhaled Ultrafine Cadmium Oxide Particles in the Rat Lung.* Inhal. Toxicol; 16(1): 83-92, (2004).
22. Kirschvink N, Martin N, Fievez L, Smith N, Marlin D. *Airway inflammation in cadmium-exposed rats is associated with pulmonary oxidative stress and emphysema.* Free Radic Res; 40: 241-50, (2006).
23. Boucher RC. *New concepts of the pathogenesis of cystic fibrosis lung disease.* Eur Respir J; 23:146-158, (2004).
24. Rennolds J, Butler S, Maloney K, Boyaka PN, Davis IC, Knoell DL, Parinandi NL, Cormet-Boyaka E. *Cadmium Regulates the Expression of the CFTR Chloride Channel in Human Airway Epithelial Cells.* Toxicol Sci. 116(1): 349-358, (2010).
25. Kundu S, Sengupta S, Chatterjee S, Mitra S, Bhattacharyya A. *Cadmium induces lung inflammation independent of lung cell proliferation: a molecular approach.* J Inflamm (Lond). 6-19, (2009).
26. Del Raso NJ, Foy BD, Gearhart JM, Frazier JM. *Cadmium Uptake Kinetics in Rat Hepatocytes: Correction for Albumin Binding.* Toxicol Sci; 72: 19-30, (2003).
27. Tracy A Chin and Douglas M Templeton. *Protective elevations of glutathione and metallothionein in cadmium-exposed mesangial cells.* Toxicology; 77:145-56, (1993).
28. Tzirogiannis CN, Panoutsopoulos GI, Demonakou MD, Hereti RI, Alexandropoulou KN, Basayannis AC, Mykoniatis MG. *Time-course of cadmium-induced acute hepatotoxicity in the rat liver: the role of apoptosis.* Arch Toxicol; 77: 694-701, (2003).
29. Dudley RE, Donald Svoboda J and Curtis Klaassen D. *Time course of cadmium-induced ultrastructural changes in rat liver.* Toxicol Appl Pharmacol; 76(1): 150-60, (1984).
30. Dudley RE, Svoboda DJ, Klaassen CD. *Acute exposure to cadmium causes severe liver injury in rats.* Toxicol Appl Pharmacol; 65:302-13, (1982).
31. Habeebu Sultan S, Jie Liu, Yaping Liu, Curtis D. Klaassen. *Metallothionein-Null Mice Are More Sensitive than Wild-Type Mice to Liver Injury Induced by Repeated Exposure to Cadmium.* Toxicol Sci; 55 (1): 223-32, (2000).
32. Goering PL, Klaassen CD. *Altered subcellular distribution of cadmium following cadmium pretreatment: possible mechanism of tolerance to*

- cadmium-induced lethality. ToxicolApplPharmacol; 70(2):195-203,(1983).*
33. Harstad Eric B and Klaassen Curtis D. *Tumor Necrosis Factor- α -Null Mice are not Resistant to Cadmium Chloride-Induced Hepatotoxicity.* *ToxicolApplPharmacol; 179(3):155-62, (2002).*
 34. Walter PC, Joshua RE, Daniel WN, James M.W, Aaron B, William D. A. *The Vascular System as a Target of Metal Toxicity.* *ToxicolSci; 102(2): 207-18,(2008).*
 35. Ana Navas A, Maria Tellez-P, Guallar E, Paul M, Ellen S, Bernard J, Virginia W. *Blood Cadmium and Lead and Chronic Kidney Disease in US Adults: A Joint Analysis.* *Am J Epidemiol; 170:1156-64,(2009).*
 36. Brzóška MM, Kamiński M, Supernak-Bobko D, Zwierz K, Moniuszko-Jakoniuk J. *Changes in the structure and function of the kidney of rats' chronically exposed to cadmium .I. Biochemical and histopathological studies.* *Arch Toxicol; 77:344-52.(2003).*
 37. Stoepler. Cadmium. In *Metals and their compounds in the environment.* Edited by E M. Weinheim, Verlag Chemie: 805-49.(1991).
 38. Klaassen Curtis D, Jie Liu, and Supratim Choudhuri. *An Intracellular Protein to Protect Against Cadmium Toxicity.* *Annu Rev Pharmacol Toxicol 39:267-94, (1999).*
 39. Hassoun Ezdihar A and Stohs Sidney J. *Cadmium-induced production of superoxide anion and nitric oxide, DNA single strand breaks and lactate dehydrogenase leakage in J774A.1 cell cultures.* *Toxicology; 112(3):219-26,(1996).*
 40. Bray TM and Bettger WJ. *The physiologic role of zinc as an antioxidant.* *Free RadicBiol Med; 8: 281- 91.(1990).*
 41. Kondoh M, Araragi S, Sato K, Higashimoto M, Takiguchi M, Sato M. *Cadmium induces apoptosis partly via caspase-9 activation in HL-60 cells.* *Toxicology; 170(1-2):111-7.(2002).*
 42. Rudolph S, Tim SN, Tom R, Lutgarde T, Dirk V, Kuznetsova T, Etienne Van Hecke, Harry AR, and Jan AS. *Bone Resorption and Environmental Exposure to Cadmium in Women: A Population Study.* *Environ Health Perspect; 116(6): 777-783.(2008).*
 43. Friberg L, Kjellstro M T, Nordberg G F. *Handbook on the toxicology of metals:* PP.130-184.
 44. Kjellström T. *Mechanism and epidemiology of bone effects of cadmium.* *IARC Sci Pub; (118):301-310.(1992)*
 45. Maryka HB. *Cadmium osteotoxicity in experimental animals: Mechanisms and relationship to human exposures.* *Toxicology and Applied Pharmacology; 238(3): 258-265, (2009).*
 46. Yamano T, Shimizu M and Noda T. *Age-Related Change in Cadmium-Induced Hepatotoxicity in Wistar Rats: Role of Kupffer Cells and Neutrophils.* *Toxicology and Applied Pharmacology; 151:9-15(1998).*
 47. Michael HC and Chedrese PJ. *Endocrine Disruption by Cadmium, a Common Environmental Toxicant with Paradoxical Effects on Reproduction.* *Exp Biol Med; 229:383-92, (2004).*
 48. Nursita Ayulungit I, Balwant Singh and Lees E. *The effects of cadmium, copper, lead, and zinc on the growth and reproduction of Proisotomaminuta Tullberg (Collembola).* *Ecotoxicology and Environmental Safety; 60(3): 306-14, (2005).*
 49. Yang JM, Arnush M, Chen QY, Wu XD, Pang B, Jiang XZ. *Cadmium-induced damage to primary cultures of rat Leydig cells.* *Reprod Toxicol ; 17(5): 553-60.(2003).*
 50. Nampoothiri LP, Agarwal A and Gupta S. *Effect of co-exposure to lead and cadmium on antioxidant status in rat ovarian granulosa cells.* *Arch Toxicol; 81(3): 145-50. (2006).*
 51. Kocak M and Akcil E. *The Effects of Chronic Cadmium Toxicity on the Hemostatic System.* *Pathophysiol Haemos Thromb; 35:411-416. (2006).*
 52. Mendiola J, Moreno JM, Roca M, Vergara-Juárez N, Martínez-García MJ, García-Sánchez A, Elvira-Rendueles B, Moreno-Grau S, López-Espín JJ, Ten J, Bernabeu R, Torres-Cantero AM. *Relationships between heavy metal concentrations in three different body fluids and male reproductive parameters: a pilot study.* *Environ Health. 19:6.(2011)*
 53. Kim K, Fujimoto VY, Parsons PJ, Steuerwald AJ, Browne RW, Bloom MS. *Recent cadmium exposure among male partners may affect oocyte fertilization during in vitro fertilization (IVF).* *J Assist Reprod Genet; 27:463-68.(2010).*



***Corresponding Author:**

Bhaskar M*

*Division of Animal Biotechnology, Department of Zoology,
Sri Venkateswara University, Tirupati - 517502, AP, India.*