Original Research Article - Pharmaceutical Sciences | Open Access | UGC Approved | MCI Approved Journal

EXTRACELLULAR SYNTHESIS OF AG NANOPARTICLES USING ESCHERICHIA COLI AND THEIR ANTIMICROBIAL EFFICACY

Rajesh Kumar Shah^{1*}, Abaide Haider¹ and Loveleena Das²

¹Dept. of Zoology, D.H.S.K College, Dibrugarh, Assam, 786001

²Gyan Vigyan Academy, Dibrugarh, Assam

*Corresponding Author Email: rajeshkumarshah39@yahoo.com

ABSTRACT

Microbe mediated synthesis of nanoparticles is gaining tremendous importance in present time due to its harmless and eco-friendly nature. The present study deals with synthesis of silver nanoparticles using the bacteria Escherichia coli extracellularly and evaluation of its antimicrobial activity. The nanoparticles were characterized by various means like UV Vis spectroscopy, TEM analysis, zeta potential analysis and FTIR analysis. The antimicrobial activity was tested by agar well diffusion method against Staphylococcus aureus (MTCC 87), Proteus mirabilis (MTCC 3310), Bacillus cereus (MTCC-1305) and Aspergillus niger (MTCC-9652). From the present study it can be concluded that the synthesized Ag nanoparticles exhibits significant antimicrobial activity against both Gram positive and Gram-negative bacteria and also against the tested fungus.

KEY WORDS

Antibacterial, Antifungal, Biosynthesis, Microbe, Silver

INTRODUCTION

Nanotechnology is a vast and emerging area that deals with the synthesis and application of nanoparticles [1]. Biosynthesis of nanoparticles are more preferred over physical and chemical method as it is eco-friendly and occur in ambient conditions [2, 3]. Microbe mediated synthesis can occur both extracellularly intracellularly [4, 5]. The former is however simple and cheap. Silver nanoparticles are considered to be unique because of its wide application in different fields like technology, agriculture, environmental technology and medicine [6, 7, 8]. Several workers have reported the therapeutic properties of Ag nanoparticles viz. antiviral [9, 10, 11, 12], antibacterial, anticancerous, anti-inflammatory [13, 14] etc. The present study aimed to synthesize Ag nanoparticles extracellularly using Escherichia coli and to evaluate its antimicrobial activity against some selected pathogenic microbes.

MATERIALS AND METHOD

Microorganisms: Microorganisms were procured from Microbial Type Culture Collection and Gene bank (IMTECH, Chandigarh, India). Ag nanoparticles were synthesized using *Escherichia coli* (MTCC 10312). The antimicrobial activity was carried out against *Staphylococcus aureus* (MTCC 87), *Proteus mirabilis* (MTCC-3310), *Bacillus cereus* (MTCC-1305) and *Aspergillus niger* (MTCC-9652).

Biosynthesis of Silver nanoparticles: Escherichia coli strain was freshly inoculated in a conical flask containing 100 ml of nutrient broth. The flask was incubated for 24 hours at 37° C. After incubation, the culture was centrifuged to remove the pellets. The supernatant was used for the synthesis of nanoparticles by mixing with AgNO₃ solution at 1 mM final concentration [15].

Characterization of Silver nanoparticles: The synthesized nanoparticle were characterized by UV Visible spectrophotometry and TEM analysis. The distribution was recorded using particle size analyzer.



FTIR analysis was performed for the detection of the organic functional groups which remain attached to the surface of nanoparticles. The stability of the synthesized nanoparticles were confirmed by zeta potential measurement [2].

Antibacterial activity:

Antimicrobial activity was tested by Agar Well Diffusion method [16]. Ampicillin was used as positive control. Diameter of the zone of inhibition was measured in cm and expressed as Mean \pm Standard Deviation.

RESULTS AND DISCUSSION

It was observed that the colour of the reaction mixture (culture supernatant incubated with $AgNO_3$) changed from yellow to brown with time. This indicated the synthesis of nanoparticles. Due to surface plasmon resonance, the silver nanoparticles are reported to produce intense absorption peak in UV absorption spectra [15]. The UV-Vis spectra analysis showed an absorption peak at 255.5 and 313 nm (Fig. 1). TEM analysis revealed the shape and size of the nanoparticles. They were found to be spherical in shape and were less than 50nm in size (Fig. 2). The size distribution of Ag nanoparticles were found to be in average 297.7 nm (Fig 3). Furthermore, the particles carried a charge of -12.6 mV. The negative zeta potential signifies that the nanoparticles were stable.

The FTIR spectra (Fig 4) shows peak at 3336.93 cm⁻¹, 1633.31 cm⁻¹, 1478.76 cm⁻¹ indicating the presence of capping agents. The sharp peak at 3336.93 cm⁻¹ is assigned for N-H stretching vibrations indicating H bonding and the presence of amines [17] and the band at 1633.31 cm⁻¹ corresponds to alkanes. The peak at 1478.76 cm⁻¹ also corresponds to amine group [18]. Microbes are considered to be suitable agents for synthesis of nanoparticles. Several workers have also successfully synthesized nanoparticles using microbes [15, 19, 20, 21, 22]. The different groups found in bacterial proteins like free amine may be responsible for formation and stabilization of Ag nanoparticles [23, 24]. Results of the antimicrobial activity of AgNPs are shown in table I. From the observations, it has been found that the synthesised AgNPs showed effective anti-microbial activity against the tested microbes such as Staphylococcus aureus, Proteus mirabilis, Bacillus cereus & Aspergillus niger. The maximum zone of inhibition has been found against Proteus mirabilis whereas the minimum against Aspergillus niger. The result is in accordance with the findings of other workers like Gurunathan et al. [25, 26]. These antimicrobial activity of Ag nanoparticles may be due to its ability to inhibit DNA replication [27]. They are also known to affect the membrane bound enzymes of the microbes [28, 29].

Table I: Antimicrobial activity of Ag nanoparticles

SI.	Microbes	Zone of inhibition (in cm)						
No.		Day 1	Day 5	Day 12	Day 18	Day 22	Day 26	Ampicillin
1	Proteus mirabilis	2.1 ± 0.50	2.3± 0.50	2.7± 0.75	2.5± 0.50	2.5± 1.00	2.9± 0.00	1.3 ± 0.50
2	Staphylococcus	1.8± 0.33	2.1± 0.50	2.0 ± 1.00	1.9± 0.50	1.9± 0.50	2.0± 1.00	2.4 ± 0.50
	aureus							
3	Bacillus cereus	1.7 ± 0.50	1.9± 0.50	1.8± 0.50	1.9± 0.50	1.9± 0.33	1.8± 1.00	2.8 ±0.50
4	Aspergillus niger	1.8 ± 0.50	1.7± 1.00	1.8± 0.50	1.8± 0.75	1.6± 1.00	1.7± 1.00	2.8 ±0.50



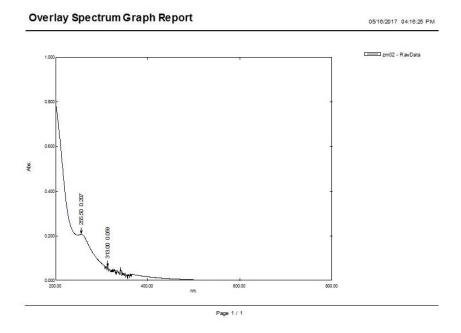
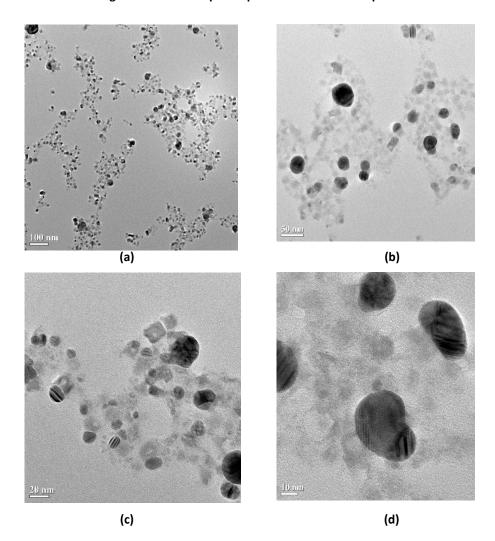
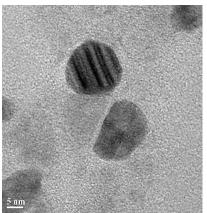
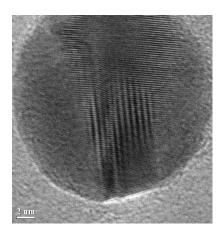


Fig 1: UV-Vis absorption spectrum of silver nanoparticles









(e) (f) Fig.2 (a-f): TEM images of Ag nanoparticles synthesized by *Escherichia coli*

Size Distribution Report by Intensity





Fig. 3: Size distribution report of Ag nanoparticles

File name: kandi colge Record Number: 4



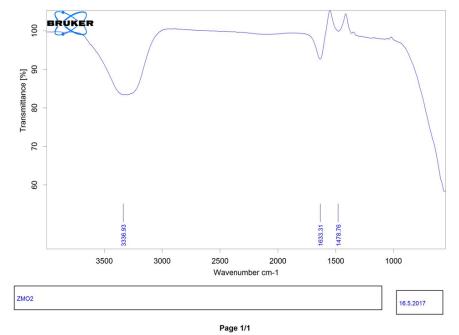


Fig 4: The FTIR spectra of the synthesized nanoparticles

CONCLUSION

The present study thus revealed that *Escherichia coli* can be effectively used to synthesize Ag nanoparticles. From the study it can also be concluded that the synthesized Ag nanoparticles exhibits significant antimicrobial activity against both Gram positive and Gram-negative bacteria and also against the tested fungus.

ACKNOWLEDGEMENT

The authors are thankful to the Institutional Level Biotech Hub of D.H.S.K College for providing the laboratory facilities to carry out the research. The authors are also thankful to Prof. S.K.Ghosh, Department of Pharmaceutical Science, Dibrugarh University, Assam for his valuable time and cooperation in UV and FT-IR analysis and Prof L.K Nath, Department of Pharmaceutical Science, Dibrugarh University, Assam for his help in Zeta Potential and Particle Size Analysis. We are also thankful to the Head of Sophisticated Analytical Instrument Facility (SAIF), NEHU, Shillong for TEM analysis.

REFERENCES

[1] Duran N., Priscyla D., Marcato PD., Alves O., De Souza G., Esposito E., Mechanistic aspects of biosynthesis of silver

- nanoparticles by several *Fusarium oxysporum* strains. *J Nanobiotechnol* 3:1–7, (2005).
- [2] Khandel P., Sahai SK., Microbes mediated synthesis of metal nanoparticles: current status and future prospects. *International Journal of Nanomaterials and Biostructures* 6(1): 1-24, (2016)
- [3] Mukherjee P., Roy M., Mandal B P., Dey G K., Mukherjee P K., Ghatak J., Tyagi A K., Kale S P., Green synthesis of highly stabilized nanocrystalline silver particles by a non-pathogenic and agriculturally important fungus T. asperellum. *Nanotechnology* 19:103–110, (2008).
- [4] Jain N., Bhargava A., Majumdar S., Tarafdar J C., Panwar J., Extracellular biosynthesis and characterization of silver nanoparticles using Aspergillus flavus NJP08: a mechanism perspective. *Nanoscale* 3:635–641, (2011).
- [5] Saifuddin N., Wong C W., NurYasumira A A., Rapid biosynthesis of silver nanoparticles using culture supernatant of bacteria with microwave irradiation. *J Chem* 6:61–70, (2009).
- [6] Galdiero S., Falanga A., Vitiello M., Cantisani M., Marra V., Galdiero M., Silver nanoparticles as potential antiviral agents. *Molecules* 16: 8894–8918, (2011).
- [7] Lu J M., Wang X., Muller C M., Wang H., Lin P H., Qizhi Y., et al. Current advances in research and clinical applications of PLGA based Nanotechnology. *Expert Rev Mol Diagn* 9: 325–341, (2009).
- [8] Sastry R K., Rashmi H B., Rao N H., Ilyas S M., Integrating nanotechnology into agri-food systems research in India: a conceptual framework. *Technol Forecast Soc* 77: 639–648, (2010).



- [9] Rogers J V., Parkinson C V., Choi Y W., Speshock J L., Hussain S M., A preliminary assessment of silver nanoparticles inhibition of monkey pox virus plaque formation. *Nanoscale Res Lett* 3: 129–133, (2008).
- [10] Baram P D., Shukla S., Gedanken A., Sarid R., Inhibition of HSV1 attachment, entry, and cell-to-cell spread by functionalized multivalent gold nanoparticles. *Small* 6: 1044–1050, (2010).
- [11] Lara H H., Ixtepan T L., Garza T E N., Rodriguez P C., PVP-coated silver nanoparticles block the transmission of cell-free and cell-associated HIV1 in human cervical culture. *J Nanobiotechnol*, 8: 15–25, (2010).
- [12] Lu L., Sun R W., Chen R., Hui C K., Ho C M., Luk J M., et al. Silver nanoparticles inhibit hepatitis B virus replication. *Antivir Ther* 13: 253–262, (2008).
- [13] Kim J S., Kuk E., Yu K N., Kim J H., Park S J., Antimicrobial effects of silver nanoparticles. *Nanomedicine* 3: 95–101, (2007).
- [14] Kuo W S., Chang C N., Chang Y T., Yeh C S., Antimicrobial gold nanorods with dual-modality photodynamic inactivation and hyperthermia. *Chem Commun*. (Camb) 32: (2009).
- [15] Das V L., Thomas R., Varghese R T., Sonia E V., Mathew J., Radhakrishnan E K., Extracellular synthesis of silver nanoparticles by the Bacillus strain CS 11 isolated from industrialized area. 3 Biotech 4:121–126, (2014)
- [16] Kelly K L., Coronado E., Zhao L L., Schatz G C., The optical properties of metal nanoparticles: the influence of size, shape and dielectric environment. *J Phys Chem B*, 107: 668–677, (2003).
- [17] Saha S., Sarkar J., Chattopadhayay D., Patra S., Chakraborty A., Acharya K., Production of silver nanoparticles by a phytopathogenic fungus Bipolaris nodulosa and its antimicrobial activity. *Dig J Nanomater Biostruct*. 5: 887–895, (2010).
- [18] Dyer J R, Applications of absorption spectroscopy of organic compounds, Prentice-Hall of India pvt. Ltd., New Delhi (1987).
- [19] Elgorban A M., Aref S M., Seham S .M, Elhindi K M., Bahkali A H., Sayed S R., Manal M A., Mycosphere Extracellular synthesis of silver nanoparticles using Aspergillus versicolor and evaluation of their activity on

- plant pathogenic fungi. *Mycosphere*, 7 (6): 844–852, (2016)
- [20] Vithiya K., Kumar R., Sen S., Bacillus Sp. mediated extracellular synthesis of silver nanoparticles *Int J Pharm Pharm Sci* 6(2): 525-527, (2014).
- [21] Elbeshehy E K F., Ahmed M E., George A., Silver nanoparticles synthesis mediated by new isolates of *Bacillus* spp., nanoparticle characterization and their activity against Bean Yellow Mosaic Virus and human pathogens. *Frontiers in Microbiology* 6(435):1-13(2015)
- [22] Paul D., Sinha S N., Extracellular Synthesis of Silver Nanoparticles Using Pseudomonas aeruginosa KUPSB12 and Its Antibacterial Activity. *Jordan J Biol Sci*, 7 (4): 245-250, (2014).
- [23] Babu M M G., Gunasekaran P., Production and structural characterization of crystalline silver nanoparticles from Bacillus cereus isolate. *Colloids Surf B*, 74:191-195, (2009).
- [24] Balaji D S., Basavaraja S., Deshpande R., Mahesh D., Prabhakar B K., Venkataraman A., Extracellular biosynthesis of functionalized silver nanoparticles by strains of *Cladosporium cladosporioides* fungus. *Colloids Surf B*, 68:88-92, (2009).
- [25] Gurunathan S., Lee K J., Kalishwaralal K., Sheikpranbabu S., Vaidyanathan R., Eom S H., Antiangiogenic properties of silver nanoparticles. *Biomaterials* 30:6341–6350, (2009a).
- [26] Gurunathan S., Kalishwaralal K., Vaidyanathan R., Venkataraman D., Pandian S R K., Muniyandi J., Hariharan N., Eom S H., Biosynthesis, purification and characterization of silver nanoparticles using *Escherichia coli. Colloids Surf B*, 74(1):328–335, (2009b)
- [27] Feng Q L., Wu J., Chen G Q., Cui F Z., Kim T N., Kim J O., A mechanistic study of the antibacterial effect of silver ions on Escherichia coli and Staphylococcus aureus. J Biomed Mater Res Part B Appl Biomater, 52: 662–668, (2000).
- [28] Bragg P D., Rainnie D J., The effect of silver ions on the respiratory chains of *Escherichia coli. Can J Microbiol* 20, 883–889, (1974).
- [29] McDonnell G., Russell A D., Antiseptics and disinfectants: activity, action, and resistance. *Clinical Microbiology Reviews* 12, 147–179, (1999).

Corresponding Author: Rajesh Kumar Shah

Email: rajeshkumarshah39@yahoo.com